

Polyphenols as Dual Modulators of Insulin Sensitivity and Lipid Metabolism in Obese Diabetic Patients

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ABSTRACT

Obesity and type 2 diabetes mellitus (T2DM) constitute two of the most pervasive and interconnected metabolic disorders worldwide. Characterized by insulin resistance and dysregulated lipid metabolism, these conditions contribute to cardiovascular disease, hepatic dysfunction, and systemic inflammation. As researchers intensify their search for natural, multi-targeted therapeutic agents, polyphenols have emerged as promising candidates. These plant-derived compounds, abundant in fruits, vegetables, teas, and spices, have demonstrated significant potential to simultaneously enhance insulin sensitivity and regulate lipid metabolism. This comprehensive review explores the molecular mechanisms by which polyphenols influence key metabolic pathways, reviews preclinical and clinical findings, and discusses the therapeutic potential, limitations, and future prospects of polyphenol-based interventions in obese diabetic patients.

Keywords: Polyphenols, Insulin Sensitivity, Lipid Metabolism, Obesity, Type 2 Diabetes Mellitus

INTRODUCTION

The global surge in obesity and type 2 diabetes mellitus (T2DM) represents a major public health concern and places a substantial burden on healthcare systems worldwide. These two metabolic disorders are closely linked, often coexisting and exacerbating one another in a vicious cycle[1–5]. Central to their pathophysiology is insulin resistance, a state in which cells fail to respond effectively to insulin, resulting in impaired glucose uptake and dysfunctional lipid metabolism, which contributes to ectopic fat deposition, chronic inflammation, and further metabolic derangement[6–8]. Traditional pharmacological interventions, such as metformin, sulfonylureas, and insulin therapy, remain the cornerstone of T2DM management. Meanwhile, weight loss drugs, including orlistat and GLP-1 receptor agonists, are often prescribed to address obesity. However, while these medications can be effective in controlling blood sugar and promoting weight loss, they are frequently accompanied by adverse side effects such as gastrointestinal disturbances, hypoglycemia, and in some cases, cardiovascular risks [9, 10]. Moreover, these treatments typically target specific pathways rather than addressing the multifactorial nature of obesity and T2DM. Given the chronic nature of these diseases and the limitations of existing drugs, there is an urgent need for safer, more holistic strategies that can target multiple metabolic pathways simultaneously[11, 12].

In this context, nutraceuticals have garnered significant attention. Among them, polyphenols a diverse group of naturally occurring compounds found abundantly in fruits, vegetables, tea, coffee, wine, and whole grains have emerged as promising agents. Polyphenols are classified into several subclasses, including flavonoids (such as quercetin, catechins, and anthocyanins), phenolic acids (like caffeic acid and ferulic acid), stilbenes (notably resveratrol), and lignans[13–15]. Their biological versatility and natural origin make them attractive candidates for long-term dietary interventions aimed at mitigating metabolic disorders. Polyphenols are known for their potent antioxidant and anti-inflammatory properties, which can counteract two key drivers of insulin resistance: oxidative stress and chronic low-grade inflammation. By neutralizing reactive oxygen species (ROS) and modulating the activity of pro-inflammatory cytokines such as TNF- α and IL-6, polyphenols help preserve insulin signaling pathways and improve glucose utilization in peripheral tissues like muscle and adipose tissue[16].

Beyond their antioxidant effects, polyphenols have been shown to directly influence several molecular pathways implicated in glucose and lipid metabolism. For instance, certain polyphenols activate AMP-activated protein kinase (AMPK), a central metabolic regulator that enhances insulin sensitivity, stimulates glucose uptake, and promotes fatty acid oxidation [16]. AMPK activation also inhibits hepatic gluconeogenesis and lipogenesis, thereby reducing fasting blood glucose and improving lipid profiles. Additionally, polyphenols can modulate peroxisome proliferator-activated receptors (PPARs), a family of nuclear receptors involved in lipid metabolism and adipogenesis. Activation of PPAR- γ by specific polyphenols enhances insulin sensitivity and reduces adipocyte inflammation, while PPAR- α activation promotes lipid catabolism [11].

Another important mechanism by which polyphenols exert metabolic benefits is through modulation of the gut microbiota. Recent research suggests that polyphenols can positively alter the composition and activity of gut microbiota, enhancing the abundance of beneficial bacteria such as *Akkermansia muciniphila* and *Bifidobacterium* spp. These microbes produce short-chain fatty acids (SCFAs) like butyrate, which have been associated with improved gut barrier function, reduced inflammation, and better glycemic control [17]. Animal and human studies further support the metabolic benefits of polyphenols. For example, supplementation with resveratrol has been linked to improved insulin sensitivity, reduced fasting glucose, and lower triglyceride levels. Similarly, green tea catechins have been shown to reduce body fat, improve lipid metabolism, and enhance glucose uptake in skeletal muscle [17]. Clinical trials with polyphenol-rich foods such as berries, cocoa, and olive oil also demonstrate modest but consistent improvements in glycemic control and markers of metabolic health in individuals with obesity or T2DM.

Despite these promising findings, challenges remain in translating the benefits of polyphenols into clinical practice. Bioavailability is a major issue, as many polyphenols are poorly absorbed, rapidly metabolized, or excreted before exerting systemic effects. Strategies to enhance bioavailability, such as nanoencapsulation, co-administration with other nutrients, or the development of synthetic analogs, are currently being explored [14, 18, 19]. Furthermore, variability in individual responses due to genetics, microbiome composition, and dietary context adds complexity to the development of standardized dosing protocols.

Polyphenols represent a compelling adjunctive approach to the management of obesity and T2DM. Their ability to target multiple pathogenic mechanisms ranging from insulin resistance and dyslipidemia to oxidative stress and inflammation offers a holistic strategy that aligns well with the complex nature of metabolic syndrome. While more research is needed to optimize their use and ensure efficacy in diverse populations, integrating polyphenol-rich foods into the diet may serve as a safe, natural, and effective component of metabolic disease management.

2. Molecular Mechanisms Underpinning Insulin Sensitivity

2.1 Antioxidant and Anti-inflammatory Effects

Oxidative stress and chronic low-grade inflammation are intricately linked to the development and progression of insulin resistance, a key feature of type 2 diabetes mellitus (T2DM) [20–22]. In individuals with obesity and T2DM, persistent hyperglycemia and elevated circulating free fatty acids stimulate the overproduction of reactive oxygen species (ROS) in metabolic tissues such as liver, muscle, and adipose tissue. These ROS, in turn, damage cellular components including lipids, proteins, and DNA, disrupting normal cell function [23, 24]. Specifically, ROS impair insulin signaling by altering critical molecules such as insulin receptor substrates (IRS), notably through oxidative modifications that interfere with their phosphorylation and downstream activity [25]. Parallel to oxidative stress, chronic inflammation further exacerbates insulin resistance. Adipose tissue in obese individuals becomes infiltrated by macrophages and other immune cells, which secrete pro-inflammatory cytokines such as tumor necrosis factor- α (TNF- α), interleukin-1 beta (IL-1 β), and interleukin-6 (IL-6) [26]. These cytokines interfere with insulin signaling by promoting serine phosphorylation of IRS proteins and activating inflammatory pathways, such as the nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) cascade. These disruptions reduce insulin sensitivity and contribute to systemic metabolic dysfunction. Polyphenols, naturally occurring phytochemicals found in fruits, vegetables, teas, and wine, have demonstrated potent antioxidant and anti-inflammatory properties. Compounds like quercetin, resveratrol, and epigallocatechin gallate (EGCG) have been shown to scavenge ROS, thereby preserving the structure and function of insulin signaling proteins [27, 28]. Additionally, polyphenols modulate inflammatory responses by suppressing cytokine production and inhibiting pro-inflammatory enzymes such as cyclooxygenase (COX) and inducible nitric oxide synthase (iNOS) [29]. These effects are particularly evident in metabolically active tissues, such as the liver and adipose tissue, where inflammation and oxidative stress are most pronounced. As a result, polyphenols help restore insulin sensitivity and protect against metabolic dysfunction.

2.2 Modulation of Insulin Signaling Cascades

Insulin signaling is a complex and tightly regulated cascade of molecular interactions that control glucose uptake, storage, and utilization [30, 31]. The pathway begins when insulin binds to its receptor (IR) on the cell surface, leading to autophosphorylation of the receptor and recruitment of insulin receptor substrates (IRS), which then activate phosphatidylinositol 3-kinase (PI3K). PI3K catalyzes the production of phosphatidylinositol

(3,4,5)-trisphosphate (PIP₃), which recruits and activates Akt, also known as protein kinase B (PKB). Akt plays a pivotal role in facilitating the translocation of glucose transporter type 4 (GLUT4) to the plasma membrane, allowing glucose to enter the cell, particularly in skeletal muscle and adipose tissue [32, 33].

In insulin-resistant states, this signaling cascade is disrupted, often due to inhibitory serine phosphorylation of IRS proteins or defects in receptor function. Polyphenols have emerged as potential modulators of this pathway. For instance, EGCG from green tea and the flavonoid myricetin have been shown to enhance insulin signaling by promoting the phosphorylation and activation of IR and Akt [34]. These effects restore GLUT4 translocation and improve glucose uptake in peripheral tissues.

Resveratrol, a stilbene found in red wine and grapes, influences insulin signaling through activation of SIRT1, a NAD⁺-dependent deacetylase. SIRT1 activation leads to deacetylation of IRS-2 and peroxisome proliferator-activated receptor gamma coactivator-1 α (PGC-1 α), which are involved in mitochondrial function and energy metabolism [35–37]. This contributes to improved insulin sensitivity and enhanced glucose homeostasis. In liver, adipose, and muscle tissues, polyphenols have demonstrated the ability to restore proper insulin signaling, thereby counteracting the molecular defects associated with insulin resistance. These effects underscore the therapeutic potential of polyphenols in managing hyperglycemia and preventing the progression of metabolic disorders such as T2DM.

2.3 Activation of AMPK Pathway

AMP-activated protein kinase (AMPK) is a master regulator of cellular energy homeostasis and plays a crucial role in maintaining metabolic balance [38]. It is activated under conditions of energy stress, such as exercise or nutrient deprivation, where the ratio of AMP to ATP increases, signaling a low-energy state. Once activated, AMPK orchestrates a metabolic shift that promotes energy-producing processes while inhibiting energy-consuming pathways [32, 39]. Key functions of AMPK include stimulating glucose uptake via GLUT4 translocation, enhancing fatty acid oxidation, and inhibiting lipid synthesis and gluconeogenesis in the liver. Polyphenols have been found to activate AMPK through various mechanisms, including the stimulation of upstream kinases such as liver kinase B1 (LKB1) and calcium/calmodulin-dependent protein kinase kinase beta (CaMKK β). Compounds like curcumin and EGCG enhance AMPK activity, thereby promoting metabolic adaptations beneficial for glucose and lipid homeostasis. For example, AMPK activation inhibits the enzyme acetyl-CoA carboxylase (ACC), leading to a decrease in malonyl-CoA levels, which relieves inhibition of carnitine palmitoyltransferase I (CPT1). This facilitates the transport of fatty acids into mitochondria for oxidation, thereby reducing lipid accumulation in tissues [40, 41].

Additionally, AMPK activation by polyphenols suppresses hepatic gluconeogenesis by downregulating key enzymes such as phosphoenolpyruvate carboxykinase (PEPCK) and glucose-6-phosphatase (G6Pase), reducing endogenous glucose production. This contributes to improved glycemic control [42, 43]. The metabolic effects of AMPK activation also extend to the enhancement of mitochondrial biogenesis and oxidative capacity, further improving insulin sensitivity. The ability of polyphenols to activate AMPK underscores their role as metabolic regulators, offering a promising avenue for therapeutic interventions targeting obesity, insulin resistance, and T2DM.

2.4 Influence on Gut Microbiota

The gut microbiota is a dynamic and diverse community of microorganisms that reside in the human gastrointestinal tract and exert profound effects on host metabolism, immune regulation, and overall health [44–46]. Emerging evidence suggests that disruptions in gut microbial composition, commonly referred to as dysbiosis, are closely associated with the development of metabolic disorders such as obesity and T2DM. In particular, dysbiosis is characterized by a reduced microbial diversity, altered Firmicutes:Bacteroidetes ratio, and a decline in beneficial microbial species [47].

Polyphenols have gained attention for their prebiotic-like effects on the gut microbiome. Due to their limited absorption in the small intestine, a significant proportion of dietary polyphenols reach the colon, where they are metabolized by gut microbes into bioactive phenolic compounds. These microbial metabolites, such as urolithins and phenyl- γ -valerolactones, possess anti-inflammatory and insulin-sensitizing properties, contributing to systemic metabolic benefits [48]. Importantly, polyphenols can selectively modulate the gut microbial ecosystem by promoting the growth of beneficial bacteria like *Akkermansia muciniphila* and *Bifidobacteria*. *Akkermansia muciniphila* is particularly noteworthy for its role in maintaining gut barrier integrity and improving insulin sensitivity. By fostering a healthier microbial environment, polyphenols indirectly reduce endotoxemia and systemic inflammation, which are key contributors to metabolic dysfunction [49].

Additionally, polyphenols influence short-chain fatty acid (SCFA) production, such as butyrate, propionate, and acetate, which have beneficial effects on glucose metabolism, lipid regulation, and immune function. These SCFAs serve as signaling molecules and energy substrates for colonocytes, further linking gut health to metabolic homeostasis [42, 48]. The interaction between polyphenols and the gut microbiome represents a promising frontier in the prevention and management of metabolic diseases, suggesting that dietary

interventions targeting the microbiota could be an effective strategy for improving insulin sensitivity and metabolic health.

3. Polyphenols and Lipid Metabolism

3.1 Enhancement of Fatty Acid Oxidation

Fatty acid oxidation is a crucial metabolic process responsible for breaking down long-chain fatty acids into acetyl-CoA, which enters the Krebs cycle for energy production. In individuals with obesity and T2DM, this process is often impaired, leading to the accumulation of lipids in non-adipose tissues such as the liver and skeletal muscle[50]. This ectopic fat deposition contributes to insulin resistance, mitochondrial dysfunction, and systemic metabolic imbalance. Enhancing fatty acid oxidation can therefore play a vital role in restoring metabolic homeostasis and preventing the progression of insulin resistance. Polyphenols such as resveratrol and chlorogenic acid have been shown to significantly enhance fatty acid oxidation through multiple molecular pathways. Resveratrol activates SIRT1, a key regulator of mitochondrial function, which in turn stimulates peroxisome proliferator-activated receptor gamma coactivator-1 α (PGC-1 α)[36, 51, 52]. PGC-1 α is a master regulator of mitochondrial biogenesis and oxidative metabolism, and its activation promotes the expression of genes involved in mitochondrial fatty acid oxidation. This leads to an increase in both the number and function of mitochondria, improving the cell's ability to oxidize fatty acids efficiently.

Chlorogenic acid, commonly found in coffee and certain fruits, enhances the expression and activity of carnitine palmitoyltransferase I (CPT1), the rate-limiting enzyme in the transport of long-chain fatty acids into mitochondria[53–55]. By facilitating this transport step, chlorogenic acid ensures that fatty acids are available for β -oxidation, thus reducing intracellular lipid accumulation. Increased fatty acid oxidation also leads to lower levels of lipotoxic intermediates like diacylglycerol and ceramides, which are known to interfere with insulin signaling. Altogether, these effects underscore the capacity of polyphenols to improve energy metabolism, reduce lipid overload in tissues, and enhance insulin sensitivity by promoting efficient fatty acid oxidation.

3.2 Inhibition of Lipogenesis

Lipogenesis, the metabolic process by which acetyl-CoA is converted into fatty acids and subsequently stored as triglycerides, is typically upregulated in states of energy excess such as obesity [56, 57]. This process is tightly regulated by transcription factors including sterol regulatory element-binding protein-1c (SREBP-1c) and carbohydrate-responsive element-binding protein (ChREBP)[4]. These transcription factors stimulate the expression of lipogenic enzymes such as fatty acid synthase (FAS) and acetyl-CoA carboxylase (ACC), driving hepatic lipid accumulation and promoting the development of non-alcoholic fatty liver disease (NAFLD), a condition strongly linked to insulin resistance and T2DM[5].

Polyphenols have been shown to exert inhibitory effects on de novo lipogenesis through multiple mechanisms. Curcumin, derived from turmeric, suppresses the expression of SREBP-1c and its downstream targets, thereby reducing the synthesis of fatty acids and triglycerides in the liver. EGCG from green tea similarly inhibits SREBP-1c as well as ChREBP, further curtailing the transcription of lipogenic enzymes[58–60]. Naringenin, a citrus flavonoid, blocks the activation of liver X receptor alpha (LXR α), a nuclear receptor that promotes lipogenesis, thereby exerting an upstream regulatory effect[61, 62]. The inhibition of these transcriptional regulators leads to a decrease in hepatic lipid synthesis and accumulation, which helps reverse steatosis and restore liver insulin sensitivity. Furthermore, the reduction in lipogenic activity limits the production of harmful lipid intermediates that impair insulin signaling pathways. These actions not only benefit hepatic metabolism but also have systemic implications for lipid and glucose homeostasis[62]. By targeting key molecular players in the lipogenic pathway, polyphenols offer a promising nutritional strategy for managing dyslipidemia, hepatic steatosis, and associated metabolic disturbances in obesity and T2DM.

3.3 Regulation of Lipid Transport and Storage

Lipid metabolism encompasses not only synthesis and oxidation but also the transport and storage of lipids within the body. Dysregulation in lipid transport can lead to elevated circulating triglycerides, abnormal lipoprotein profiles, and lipid deposition in non-adipose tissues—all of which are associated with increased cardiometabolic risk[63]. Polyphenols play a significant role in modulating these aspects of lipid handling, thereby contributing to overall metabolic health. One important mechanism involves the regulation of lipoprotein metabolism[16, 40]. Polyphenols enhance the activity of lipoprotein lipase (LPL), an enzyme responsible for hydrolyzing triglycerides in circulating chylomicrons and very low-density lipoproteins (VLDL), facilitating the uptake of free fatty acids by peripheral tissues[64]. Increased LPL activity leads to a more efficient clearance of triglyceride-rich lipoproteins from the bloodstream, resulting in improved plasma lipid profiles. Additionally, polyphenols have been shown to suppress the activity of microsomal triglyceride transfer protein (MTP), which is crucial for VLDL assembly and secretion in the liver. By inhibiting MTP, polyphenols reduce hepatic triglyceride export, thereby lowering plasma VLDL and LDL levels[64].

In adipose tissue, polyphenols influence lipid storage by modulating the expression of proteins involved in lipid droplet formation and breakdown. Apigenin and luteolin, for instance, regulate the expression of perilipin, a key protein that coats lipid droplets and controls lipolysis. Proper regulation of perilipin ensures that lipolysis occurs

in a controlled manner, preventing excessive release of free fatty acids into the circulation, which could otherwise contribute to insulin resistance[64].

By improving both lipid transport and storage mechanisms, polyphenols help maintain lipid balance and prevent ectopic fat deposition. These regulatory effects reduce cardiovascular risk and improve insulin sensitivity, making polyphenols valuable adjuncts in managing lipid metabolism in obesity and T2DM.

3.4 Clinical Impacts on Lipid Profiles

The lipid-modulating effects of polyphenols observed in cellular and animal models are supported by an increasing body of evidence from human clinical studies. These studies demonstrate that regular consumption of polyphenol-rich foods and supplements is associated with significant improvements in lipid profiles, which in turn may contribute to reduced cardiovascular risk in individuals with obesity and T2DM[16, 40, 65].

For instance, moderate consumption of red wine, which contains resveratrol and other polyphenols, has been linked to reductions in total cholesterol, low-density lipoprotein cholesterol (LDL-C), and triglycerides, along with an increase in high-density lipoprotein cholesterol (HDL-C)[66]. Similar effects have been observed with green tea polyphenols, particularly EGCG, which lowers plasma triglyceride and LDL-C levels while boosting HDL-C. These improvements are thought to result from the polyphenols' ability to enhance lipid clearance, reduce hepatic lipogenesis, and inhibit lipid peroxidation, thereby improving both lipid metabolism and vascular function[66].

Clinical trials have also examined polyphenol supplementation in capsule or extract form. In one trial, curcumin supplementation over eight weeks resulted in decreased triglyceride levels and improved total cholesterol-to-HDL ratios in obese individuals[67]. Another study involving quercetin supplementation showed reductions in LDL-C and improvements in markers of oxidative stress and inflammation, which are often elevated in dyslipidemic individuals[68].

While individual responses may vary, the cumulative clinical data indicate a consistent trend: polyphenols improve lipid profiles through a combination of mechanisms, including enhanced lipid clearance, reduced lipid synthesis, and improved antioxidant status[68]. These effects are particularly relevant in metabolic diseases, where dyslipidemia is a common and dangerous comorbidity. As part of a balanced diet or in supplement form, polyphenols offer a safe and natural approach to managing blood lipid levels and reducing long-term cardiovascular risk.

4. Challenges in polyphenols.

4.1 Low Bioavailability as a Limiting Factor

One of the major limitations in realizing the therapeutic potential of polyphenols is their inherently low bioavailability[18, 49]. Many polyphenolic compounds, including resveratrol, curcumin, and quercetin, exhibit poor solubility, rapid metabolism, and limited absorption in the gastrointestinal tract. After oral ingestion, polyphenols undergo extensive first-pass metabolism in the liver and intestine, resulting in glucuronidation, sulfation, or methylation that drastically reduces the concentration of active forms reaching systemic circulation. For instance, resveratrol, despite showing remarkable efficacy in preclinical studies, exhibits rapid clearance and low plasma levels in human trials, which likely attenuates its biological impact[69]. Curcumin faces similar challenges due to its low water solubility, poor intestinal absorption, and instability in neutral and basic pH conditions. These pharmacokinetic constraints pose significant barriers to translating in vitro and animal model findings into clinical benefits.

Furthermore, the variability in absorption and metabolism across individuals adds to the complexity. Factors such as gut microbiota composition, genetic differences in metabolizing enzymes, and dietary matrix influence the extent and rate at which polyphenols are absorbed and metabolized[69]. Consequently, even when high doses are consumed, only a fraction of the active compounds may exert meaningful biological effects.

These issues have prompted researchers to explore methods to enhance polyphenol bioavailability, such as structural modification, co-administration with absorption enhancers, and advanced delivery systems. Nonetheless, the inconsistency between promising laboratory results and modest clinical outcomes is often attributable to bioavailability limitations. Overcoming these hurdles remains a critical step in harnessing the full therapeutic potential of polyphenols. Without addressing this fundamental issue, the clinical application of polyphenols as metabolic modulators will remain constrained despite their extensive biological promise.

4.2 Novel Delivery Systems

In response to the bioavailability challenges associated with polyphenols, researchers have developed innovative delivery systems designed to enhance their absorption, stability, and bioactivity. These novel strategies aim to improve the therapeutic efficacy of polyphenols by increasing their solubility, protecting them from degradation, and facilitating sustained release into the bloodstream.

Nanotechnology has emerged as a promising solution. Nanoparticles, including nanoemulsions, solid lipid nanoparticles, and polymer-based nanocarriers, can encapsulate polyphenols and shield them from enzymatic degradation in the gastrointestinal tract[70–72]. These delivery systems increase intestinal permeability, prolong circulation time, and enable targeted delivery to specific tissues. For example, curcumin-loaded

nanoparticles have demonstrated superior bioavailability and anti-inflammatory effects compared to conventional curcumin supplements [73, 74]. Liposomes, which are phospholipid-based vesicles, offer another effective strategy. They enhance solubility and promote cellular uptake of hydrophobic polyphenols like resveratrol and quercetin [75–77]. Similarly, phospholipid complexes, such as phytosomes, bind polyphenols to phosphatidylcholine, facilitating improved intestinal absorption and liver targeting. The use of bioenhancers such as piperine, derived from black pepper, has also gained attention [78]. Piperine inhibits hepatic and intestinal glucuronidation enzymes, significantly increasing the systemic availability of curcumin—by up to 2,000% in some studies. This approach allows for lower doses to be used while maintaining efficacy, reducing the risk of side effects [79].

Overall, these advanced formulation techniques represent a critical advancement in overcoming the pharmacokinetic limitations of polyphenols. By improving bioavailability and stability, they expand the clinical applicability of polyphenol-based therapies in managing metabolic diseases. However, regulatory approval, cost-effectiveness, and long-term safety of these delivery systems must also be thoroughly evaluated before widespread clinical implementation.

4.3 Safety Considerations

While polyphenols are generally regarded as safe when consumed as part of a balanced diet, their use in concentrated supplement form introduces potential safety concerns that must be carefully evaluated, especially in clinical and long-term settings. Naturally occurring polyphenols in foods like berries, tea, and vegetables have a long history of safe consumption. However, when these compounds are extracted, purified, and administered at high doses, their pharmacological activity can lead to unintended physiological effects or interactions with medications [80].

For instance, green tea catechins such as epigallocatechin gallate (EGCG), when taken in large quantities, have been associated with liver toxicity in some individuals. Cases of hepatotoxicity have been reported with high-dose green tea extract supplements, prompting health authorities to issue cautionary guidelines [29, 81]. The European Food Safety Authority (EFSA) has suggested that daily EGCG intakes above 800 mg may carry a risk of liver injury. Similarly, while curcumin is widely celebrated for its anti-inflammatory effects, high doses can cause gastrointestinal symptoms such as bloating, diarrhea, and nausea, and may interfere with iron absorption or certain drugs, including anticoagulants.

Resveratrol, though promising in metabolic health, also poses risks when consumed at pharmacological doses [36, 51, 69]. It can influence the activity of cytochrome P450 enzymes, thereby affecting the metabolism of drugs such as statins, anticoagulants, and NSAIDs. These interactions may either reduce drug efficacy or increase the risk of adverse events. Moreover, novel polyphenol delivery systems, including nanoparticles, liposomes, and phytosomes, while improving bioavailability, raise new concerns regarding long-term biocompatibility, tissue accumulation, and immunogenicity. As these technologies become more common, rigorous safety assessments, including toxicology studies, will be essential [37].

Therefore, while dietary intake of polyphenols remains safe for the general population, high-dose supplementation should be approached with caution, particularly in individuals with underlying health conditions or those taking multiple medications. Medical supervision and further long-term clinical studies are warranted to fully establish safety profiles.

5. Synergistic Effects and Dietary Integration

5.1 Synergy Among Polyphenols: Combining multiple polyphenols may enhance their efficacy through additive or synergistic mechanisms. For example, combining resveratrol and quercetin may produce more potent anti-inflammatory and insulin-sensitizing effects than either compound alone. Such combinations can target multiple pathways simultaneously, offering a holistic therapeutic approach [82].

5.2 Whole Foods vs. Supplements: Dietary patterns rich in polyphenol-containing foods (e.g., Mediterranean diet) are associated with lower incidence of T2DM and cardiovascular disease. Whole foods provide a matrix of nutrients and fiber that may enhance polyphenol effects. While supplements offer concentrated doses, they lack the complexity of whole-food sources and may not replicate the same health benefits. [83]

5.3 Integration into Lifestyle Interventions: Polyphenol intake should be incorporated within broader lifestyle modification programs, including physical activity and dietary regulation. This integrative approach can maximize metabolic benefits and improve patient adherence. Education and public health initiatives can promote consumption of polyphenol-rich foods in at-risk populations [84].

6. Future Perspectives

Future research should focus on:

- Long-term, large-scale randomized controlled trials to validate efficacy and safety.
- Personalized nutrition approaches to identify responders and optimize interventions.
- Exploration of polyphenol-microbiota interactions and their role in metabolic regulation.
- Development of bioavailable, stable, and safe polyphenol formulations for clinical use.

- Integration of polyphenols into clinical guidelines for metabolic disease management.

CONCLUSION

Polyphenols hold significant promise as dual modulators of insulin sensitivity and lipid metabolism, offering a natural, multifaceted approach to managing obesity and T2DM. Their ability to target multiple pathogenic mechanisms simultaneously makes them attractive therapeutic agents. While evidence from animal and early human studies is encouraging, more rigorous clinical trials and improved delivery systems are necessary to fully harness their potential. Incorporating polyphenol strategies into dietary and lifestyle interventions could provide a sustainable and effective means to combat the growing epidemic of metabolic diseases.

REFERENCES

1. Allocca, S., Monda, A., Messina, A., Casillo, M., Sapuppo, W., Monda, V., Polito, R., Di Maio, G., Monda, M., La Marra, M.: Endocrine and Metabolic Mechanisms Linking Obesity to Type 2 Diabetes: Implications for Targeted Therapy. *Healthcare*. 13, 1437 (2025). <https://doi.org/10.3390/healthcare13121437>
2. Chandrasekaran, P., Weiskirchen, R.: The Role of Obesity in Type 2 Diabetes Mellitus—An Overview. *International Journal of Molecular Sciences*. 25, 1882 (2024). <https://doi.org/10.3390/ijms25031882>
3. Fu, L., Baranova, A., Cao, H., Zhang, F.: Gut microbiome links obesity to type 2 diabetes: insights from Mendelian randomization. *BMC Microbiology*. 25, 253 (2025). <https://doi.org/10.1186/s12866-025-03968-8>
4. Umoru, G.U., Atangwho, I.J., David-Oku, E., Uti, D.E., De Campos, O.C., Udeozor, P.A., Nfona, S.O., Lawal, B., Alum, E.U.: Modulation of Lipogenesis by Tetracarpidium conophorum Nuts via SREBP-1/ACCA-1/FASN Inhibition in Monosodium-Glutamate-Induced Obesity in Rats. *Natural Product Communications*. 20, 1934578X251344035 (2025). <https://doi.org/10.1177/1934578X251344035>
5. David-Oku, E., Agwupuye, E.I., Obeten, U.N., Maitra, S., Subramaniyan, V., Wong, L.S., Aljarba, N.H., Kumarasamy, V.: Tetracarpidium conophorum nuts (African walnuts) up-regulated adiponectin and PPAR- γ expressions with reciprocal suppression of TNF- α gene in obesity. *J Cell Mol Med*. 28, e70086 (2024). <https://doi.org/10.1111/jcmm.70086>
6. Holmes, A., Coppey, L.J., Davidson, E.P., Yorek, M.A.: Rat Models of Diet-Induced Obesity and High Fat/Low Dose Streptozotocin Type 2 Diabetes: Effect of Reversal of High Fat Diet Compared to Treatment with Enalapril or Menhaden Oil on Glucose Utilization and Neuropathic Endpoints. *J Diabetes Res*. 2015, 307285 (2015). <https://doi.org/10.1155/2015/307285>
7. Ejemot-Nwadiaro, R.I., Betiang, P.A., Basajja, M., Uti, D.E.: Obesity and Climate Change: A Two-way Street with Global Health Implications. *Obesity Medicine*. 56, 100623 (2025). <https://doi.org/10.1016/j.obmed.2025.100623>
8. Yashi, K., Daley, S.F.: Obesity and Type 2 Diabetes. In: *StatPearls*. StatPearls Publishing, Treasure Island (FL) (2025)
9. Alum, E.U.: Optimizing patient education for sustainable self-management in type 2 diabetes. *Discov Public Health*. 22, 44 (2025). <https://doi.org/10.1186/s12982-025-00445-5>
10. Ikpozu, E.N., Offor, C.E., Igwenyi, I.O., Obaroh, I.O., Ibiyam, U.A., Ukaidi, C.U.A.: RNA-based diagnostic innovations: A new frontier in diabetes diagnosis and management. *Diabetes & Vascular Disease Research*. 22, 14791641251334726 (2025). <https://doi.org/10.1177/14791641251334726>
11. Uti, D.E., Atangwho, I.J., Alum, E.U., Egba, S.I., Ugwu, O.P.-C., Ikechukwu, G.C.: Natural Antidiabetic Agents: Current Evidence and Development Pathways from Medicinal Plants to Clinical use. *Natural Product Communications*. 20, 1934578X251323393 (2025). <https://doi.org/10.1177/1934578X251323393>
12. Uti, D.E., Atangwho, I.J., Omang, W.A., Alum, E.U., Obeten, U.N., Udeozor, P.A., Agada, S.A., Bawa, I., Ogbu, C.O.: Cytokines as key players in obesity low grade inflammation and related complications. *Obesity Medicine*. 54, 100585 (2025). <https://doi.org/10.1016/j.obmed.2025.100585>
13. Bakun, P., Mlynarczyk, D.T., Koczorowski, T., Cerbin-Koczorowska, M., Piwowarczyk, L., Kolasiński, E., Stawny, M., Kuźmińska, J., Jelińska, A., Goslinski, T.: Tea-break with epigallocatechin gallate derivatives – Powerful polyphenols of great potential for medicine. *European Journal of Medicinal Chemistry*. 261, 115820 (2023). <https://doi.org/10.1016/j.ejmech.2023.115820>
14. Ciupeî, D., Colişar, A., Leopold, L., Stănilă, A., Diaconeasa, Z.M.: Polyphenols: From Classification to Therapeutic Potential and Bioavailability. *Foods*. 13, 4131 (2024). <https://doi.org/10.3390/foods13244131>
15. Nagahawatta, D.P., Liyanage, N.M., Jayawardena, T.U., Jeon, Y.-J.: Marine Polyphenols in Cardiovascular Health: Unraveling Structure–Activity Relationships, Mechanisms, and Therapeutic Implications. *Int J Mol Sci*. 25, 8419 (2024). <https://doi.org/10.3390/ijms25158419>

16. Nowak, M., Tryniszewski, W., Sarniak, A., Wlodarczyk, A., Nowak, P.J., Nowak, D.: Concentration Dependence of Anti- and Pro-Oxidant Activity of Polyphenols as Evaluated with a Light-Emitting Fe²⁺-Egta-H₂O₂ System. *Molecules*. **27**, 3453 (2022). <https://doi.org/10.3390/molecules27113453>
17. Temis-Cortina, J.A., Prada-Ramírez, H.A., Ríos-Guerra, H., Espinosa-Raya, J., Gómez-Pliego, R.: Response of *Akkermansia muciniphila* to Bioactive Compounds: Effects on Its Abundance and Activity. *Fermentation*. **11**, 427 (2025). <https://doi.org/10.3390/fermentation11080427>
18. Aqil, F., Munagala, R., Jeyabalan, J., Vadhanam, M.V.: Bioavailability of phytochemicals and its enhancement by drug delivery systems. *Cancer Lett.* **334**, 133–141 (2013). <https://doi.org/10.1016/j.canlet.2013.02.032>
19. Alum, E.U.: Role of phytochemicals in cardiovascular disease management: Insights into mechanisms, efficacy, and clinical application. *Phytomedicine Plus*. **5**, 100695 (2025). <https://doi.org/10.1016/j.phyplu.2024.100695>
20. Uti, D.E. & Offor, C.E.: Redox Signaling Disruption and Antioxidants in Toxicology: From Precision Therapy to Potential Hazards. *Cell Biochemistry and Biophysics*. (2025). <https://doi.org/10.1007/s12013-025-01846-8>
21. Chrastina, M., Poništ, S., Tóth, J., Czigle, S., Pašková, L., Vyletelová, V., Švík, K., Bauerová, K.: Combination Therapy of Carnosic Acid and Methotrexate Effectively Suppressed the Inflammatory Markers and Oxidative Stress in Experimental Arthritis. *Molecules*. **27**, 7115 (2022). <https://doi.org/10.3390/molecules27207115>
22. Ibiam, U.A., Uti, D.E., Ejeogo, C.C., Orji, O.U., Aja, P.M., Nwamaka, E.N., Alum, E.U., Chukwu, C., Aloke, C., Itodo, M.O., Agada, S.A., Umoru, G.U., Obeten, U.N., Nwobodo, V.O.G., Nwadium, S.K., Udoudoh, M.P.: *Xylopiya aethiopic*a Attenuates Oxidative Stress and Hepatorenal Damage in Testosterone Propionate-Induced Benign Prostatic Hyperplasia in Rats. *Journal of Health and Allied Sciences NU*. **14**, 477–485 (2024). <https://doi.org/10.1055/s-0043-1777836>
23. Umoru, G.U., Uti, D.E., Aja, P.M., Ugwu, O.P., Orji, O.U., Nwali, B.U., Ezeani, N.N., Edwin, N., Orinya, F.O.: Hepato-protective effect of ethanol leaf extract of *Datura stramonium* in alloxan-induced diabetic albino rats. *Journal of Chemical Society of Nigeria*. **47**, (2022). <https://doi.org/10.46602/jcsn.v47i5.819>
24. Krishnamoorthy, R., Gatasheh, M.K., Subbarayan, S., Vijayalakshmi, P., Uti, D.E.: Protective Role of Jimson Weed in Mitigating Dyslipidemia, Cardiovascular, and Renal Dysfunction in Diabetic Rat Models: In Vivo and in Silico Evidence. *Natural Product Communications*. **19**, 1934578X241299279 (2024). <https://doi.org/10.1177/1934578X241299279>
25. Udeozor, P.A., Ibiam, U.A., Uti, D.E., Umoru, G.U., Onwe, E.N., Mbonu, F.O., Omang, W.A., Ijoganu, S.I., Anaga, C.O., Mbah, J.O., Nwadium, S.K.: Antioxidant and Anti-Anemic Effects of Ethanol Leaf Extracts of *Mucuna poggei* and *Telfairia occidentalis* in Phenyl-Hydrazine-Induced Anemia in Wistar Albino Rats. *Ibnosina Journal of Medicine and Biomedical Sciences*. **14**, 116–126 (2022). <https://doi.org/10.1055/s-0042-1756684>
26. Alharbi, H.O.A., Alshebremi, M., Babiker, A.Y., Rahmani, A.H.: The Role of Quercetin, a Flavonoid in the Management of Pathogenesis Through Regulation of Oxidative Stress, Inflammation, and Biological Activities. *Biomolecules*. **15**, 151 (2025). <https://doi.org/10.3390/biom15010151>
27. Alum, E.U.: Climate change and its impact on the bioactive compound profile of medicinal plants: implications for global health. *Plant Signal Behav.* **19**, 2419683 (2024). <https://doi.org/10.1080/15592324.2024.2419683>
28. Saeed, E., Javed, F., Rana, Z., Perveen, R., Mallhi, I.Y., Amjad, I., Maqsood, Q., Chaudhary, N.A., Tahir, S.B., Fatima, A., Rasheed, N.F.: Bioactive Compounds, Their Mechanisms of Action, and Cardioprotective Effects of Pomegranate (*Punica granatum*): A Comprehensive Review. *eFood*. **6**, e70075 (2025). <https://doi.org/10.1002/efd2.70075>
29. Jiang, P., Xu, C., Chen, L., Chen, A., Wu, X., Zhou, M., Haq, I.U., Mariyam, Z., Feng, Q.: EGCG inhibits CSC-like properties through targeting miR-485/CD44 axis in A549-cisplatin resistant cells. *Mol Carcinog*. **57**, 1835–1844 (2018). <https://doi.org/10.1002/mc.22901>
30. Baxter, R.C.: Signaling Pathways of the Insulin-like Growth Factor Binding Proteins. *Endocrine Reviews*. **44**, 753–778 (2023). <https://doi.org/10.1210/endrev/bnad008>
31. Hua, H., Kong, Q., Yin, J., Zhang, J., Jiang, Y.: Insulin-like growth factor receptor signaling in tumorigenesis and drug resistance: a challenge for cancer therapy. *J Hematol Oncol*. **13**, 64 (2020). <https://doi.org/10.1186/s13045-020-00904-3>
32. Sayem, A.S.M., Arya, A., Karimian, H., Krishnasamy, N., Ashok Hasannis, A., Hossain, C.F.: Action of Phytochemicals on Insulin Signaling Pathways Accelerating Glucose Transporter (GLUT4) Protein Translocation. *Molecules*. **23**, 258 (2018). <https://doi.org/10.3390/molecules23020258>
33. Vella, V., De Francesco, E.M., Lappano, R., Muoio, M.G., Manzella, L., Maggiolini, M., Belfiore, A.: Microenvironmental Determinants of Breast Cancer Metastasis: Focus on the Crucial Interplay Between

- Estrogen and Insulin/Insulin-Like Growth Factor Signaling. *Front Cell Dev Biol.* 8, 608412 (2020). <https://doi.org/10.3389/fcell.2020.608412>
34. Le, T.K.C., Dao, X.D., Nguyen, D.V., Luu, D.H., Bui, T.M.H., Le, T.H., Nguyen, H.T., Le, T.N., Hosaka, T., Nguyen, T.T.T.: Insulin signaling and its application. *Front. Endocrinol.* 14, (2023). <https://doi.org/10.3389/fendo.2023.1226655>
 35. Cavalcante de Freitas, P.G., Rodrigues Arruda, B., Araújo Mendes, M.G., Barroso de Freitas, J.V., da Silva, M.E., Sampaio, T.L., Petrilli, R., Eloy, J.O.: Resveratrol-Loaded Polymeric Nanoparticles: The Effects of D- α -Tocopheryl Polyethylene Glycol 1000 Succinate (TPGS) on Physicochemical and Biological Properties against Breast Cancer In Vitro and In Vivo. *Cancers (Basel).* 15, 2802 (2023). <https://doi.org/10.3390/cancers15102802>
 36. Čučuz, V., Cvejić, J., Gojković-Bukarica, L.: Clinical trials of resveratrol efficacy and safety. *Vojnosanitetski pregljed.* 79, 613–618 (2022)
 37. Li, Z., Zhang, Z., Ke, L., Sun, Y., Li, W., Feng, X., Zhu, W., Chen, S.: Resveratrol promotes white adipocytes browning and improves metabolic disorders in Sirt1-dependent manner in mice. *FASEB J.* 34, 4527–4539 (2020). <https://doi.org/10.1096/fj.201902222R>
 38. Srivastava, R.A.K., Pinkosky, S.L., Filippov, S., Hanselman, J.C., Cramer, C.T., Newton, R.S.: AMP-activated protein kinase: an emerging drug target to regulate imbalances in lipid and carbohydrate metabolism to treat cardio-metabolic diseases. *J Lipid Res.* 53, 2490–2514 (2012). <https://doi.org/10.1194/jlr.R025882>
 39. Park, Y., Jeong, E.M.: Glutathione Dynamics in the Tumor Microenvironment: A Potential Target of Cancer Stem Cells and T Cells. *Int J Stem Cells.* 17, 270–283 (2024). <https://doi.org/10.15283/ijsc24060>
 40. Bešlo, D., Golubić, N., Rastija, V., Agić, D., Karnaš, M., Šubarić, D., Lučić, B.: Antioxidant Activity, Metabolism, and Bioavailability of Polyphenols in the Diet of Animals. *Antioxidants (Basel).* 12, 1141 (2023). <https://doi.org/10.3390/antiox12061141>
 41. Rudrapal, M., Khairnar, S.J., Khan, J., Dukhyil, A.B., Ansari, M.A., Alomary, M.N., Alshabrimi, F.M., Palai, S., Deb, P.K., Devi, R.: Dietary Polyphenols and Their Role in Oxidative Stress-Induced Human Diseases: Insights Into Protective Effects, Antioxidant Potentials and Mechanism(s) of Action. *Front Pharmacol.* 13, 806470 (2022). <https://doi.org/10.3389/fphar.2022.806470>
 42. Okpoghono, J., Isoje, E.F., Igbuku, U.A., Ekayoda, O., Omoike, G.O., Adonor, T.O., Igue, U.B., Okom, S.U., Ovowa, F.O., Stephen-Onojedje, Q.O., Ejueyitsi, E.O., Seigha, A.A.: Natural polyphenols: A protective approach to reduce colorectal cancer. *Heliyon.* 10, e32390 (2024). <https://doi.org/10.1016/j.heliyon.2024.e32390>
 43. Plamada, D., Vodnar, D.C.: Polyphenols—Gut Microbiota Interrelationship: A Transition to a New Generation of Prebiotics. *Nutrients.* 14, 137 (2021). <https://doi.org/10.3390/nu14010137>
 44. Ugwu, O.P.-C., Uti, D.E., Edeh, F.O., Ainebyoona, C.: Unveiling the microbial orchestra: exploring the role of microbiota in cancer development and treatment. *Discov Onc.* 16, 646 (2025). <https://doi.org/10.1007/s12672-025-02352-2>
 45. Cai, J., Sun, L., Gonzalez, F.J.: Gut microbiota-derived bile acids in intestinal immunity, inflammation, and tumorigenesis. *Cell Host Microbe.* 30, 289–300 (2022). <https://doi.org/10.1016/j.chom.2022.02.004>
 46. Ugwu, O.P.-C., Okon, M.B., Alum, E.U., Ugwu, C.N., Anyanwu, E.G., Mariam, B., Ogenyi, F.C., Eze, V.H.U., Anyanwu, C.N., Ezeonwumelu, J.O.C., Egba, S.I., Uti, D.E., Onohuean, H., Aja, P.M., Ugwu, M.N.: Unveiling the therapeutic potential of the gut microbiota–brain axis: Novel insights and clinical applications in neurological disorders. *Medicine.* 104, e43542 (2025). <https://doi.org/10.1097/MD.0000000000043542>
 47. Ugwu, O.P.-C., Alum, E.U., Okon, M.B., Obeagu, E.I.: Mechanisms of microbiota modulation: Implications for health, disease, and therapeutic interventions. *Medicine.* 103, e38088 (2024). <https://doi.org/10.1097/MD.0000000000038088>
 48. Catalkaya, G., Venema, K., Lucini, L., Rocchetti, G., Delmas, D., Daglia, M., De Filippis, A., Xiao, H., Quiles, J.L., Xiao, J., Capanoglu, E.: Interaction of dietary polyphenols and gut microbiota: Microbial metabolism of polyphenols, influence on the gut microbiota, and implications on host health. *Food Frontiers.* 1, 109–133 (2020). <https://doi.org/10.1002/fft2.25>
 49. Bié, J., Sepodes, B., Fernandes, P.C.B., Ribeiro, M.H.L.: Polyphenols in Health and Disease: Gut Microbiota, Bioaccessibility, and Bioavailability. *Compounds.* 3, 40–72 (2023). <https://doi.org/10.3390/compounds3010005>
 50. Ma, Y., Temkin, S.M., Hawkridge, A.M., Guo, C., Wang, W., Wang, X.-Y., Fang, X.: Fatty acid oxidation: An emerging facet of metabolic transformation in cancer. *Cancer Lett.* 435, 92–100 (2018). <https://doi.org/10.1016/j.canlet.2018.08.006>

51. Sikur, N., Böröczky, C., Paszternák, A., Gyöngyössi, R., Szökő, É., Varga, K., Tábi, T.: Resveratrol and Its Derivatives Diminish Lipid Accumulation in Adipocytes In Vitro—Mechanism of Action and Structure–Activity Relationship. *Nutrients*. 16, 3869 (2024). <https://doi.org/10.3390/nu16223869>
52. Zambrano, A., Molt, M., Uribe, E., Salas, M.: Glut 1 in Cancer Cells and the Inhibitory Action of Resveratrol as A Potential Therapeutic Strategy. *Int J Mol Sci*. 20, 3374 (2019). <https://doi.org/10.3390/ijms20133374>
53. Kanchanasurakit, S., Saokaew, S., Phisalprapa, P., Duangjai, A.: Chlorogenic acid in green bean coffee on body weight: a systematic review and meta-analysis of randomized controlled trials. *Systematic Reviews*. 12, 163 (2023). <https://doi.org/10.1186/s13643-023-02311-4>
54. Nguyen, V., Taine, E.G., Meng, D., Cui, T., Tan, W.: Chlorogenic Acid: A Systematic Review on the Biological Functions, Mechanistic Actions, and Therapeutic Potentials. *Nutrients*. 16, 924 (2024). <https://doi.org/10.3390/nu16070924>
55. Wang, L., Pan, X., Jiang, L., Chu, Y., Gao, S., Jiang, X., Zhang, Y., Chen, Y., Luo, S., Peng, C.: The Biological Activity Mechanism of Chlorogenic Acid and Its Applications in Food Industry: A Review. *Front. Nutr*. 9, (2022). <https://doi.org/10.3389/fnut.2022.943911>
56. Alum, E.U.: Metabolic memory in obesity: Can early-life interventions reverse lifelong risks? *Obesity Medicine*. 55, 100610 (2025). <https://doi.org/10.1016/j.obmed.2025.100610>
57. Obasi, D.C., Abba, J.N., Aniokete, U.C., Okoroh, P.N., Akwari, A.Ak.: Evolving Paradigms in Nutrition Therapy for Diabetes: From Carbohydrate Counting to Precision Diets. *Obesity Medicine*. 100622 (2025). <https://doi.org/10.1016/j.obmed.2025.100622>
58. Alam, M.S., Anwar, M.J., Maity, M.K., Azam, F., Jaremko, M., Emwas, A.-H.: The Dynamic Role of Curcumin in Mitigating Human Illnesses: Recent Advances in Therapeutic Applications. *Pharmaceuticals*. 17, 1674 (2024). <https://doi.org/10.3390/ph17121674>
59. Allahyari, H., Shamsini, L., Zamani, H.: Dual encapsulation of curcumin and ciprofloxacin in chitosan nanoparticles attenuates *Pseudomonas aeruginosa* virulence, elastinolytic potential and quorum sensing genes. *Microb Pathog*. 202, 107438 (2025). <https://doi.org/10.1016/j.micpath.2025.107438>
60. Dai, C., Lin, J., Li, H., Shen, Z., Wang, Y., Velkov, T., Shen, J.: The Natural Product Curcumin as an Antibacterial Agent: Current Achievements and Problems. *Antioxidants (Basel)*. 11, 459 (2022). <https://doi.org/10.3390/antiox11030459>
61. Hernando-Amado, S., Alcalde-Rico, M., Gil-Gil, T., Valverde, J.R., Martínez, J.L.: Naringenin Inhibition of the *Pseudomonas aeruginosa* Quorum Sensing Response Is Based on Its Time-Dependent Competition With N-(3-Oxo-dodecanoyl)-L-homoserine Lactone for LasR Binding. *Front Mol Biosci*. 7, 25 (2020). <https://doi.org/10.3389/fmolb.2020.00025>
62. Vásquez-Reyes, S., Bernal-Gámez, M., Domínguez-Chávez, J., Mondragón-Vásquez, K., Sánchez-Tapia, M., Ordaz, G., Granados-Portillo, O., Coutiño-Hernández, D., Barrera-Gómez, P., Torres, N., Tovar, A.R.: The Effects of Novel Co-Amorphous Naringenin and Fisetin Compounds on a Diet-Induced Obesity Murine Model. *Nutrients*. 16, 4425 (2024). <https://doi.org/10.3390/nu16244425>
63. Dakal, T.C., Xiao, F., Bhusal, C.K., Sabapathy, P.C., Segal, R., Chen, J., Bai, X.: Lipids dysregulation in diseases: core concepts, targets and treatment strategies. *Lipids Health Dis*. 24, 61 (2025). <https://doi.org/10.1186/s12944-024-02425-1>
64. Kumari, A., Kristensen, K.K., Ploug, M., Winther, A.-M.L.: The Importance of Lipoprotein Lipase Regulation in Atherosclerosis. *Biomedicines*. 9, 782 (2021). <https://doi.org/10.3390/biomedicines9070782>
65. Ma, J., Zheng, Y., Tang, W., Yan, W., Nie, H., Fang, J., Liu, G.: Dietary polyphenols in lipid metabolism: A role of gut microbiome. *Anim Nutr*. 6, 404–409 (2020). <https://doi.org/10.1016/j.aninu.2020.08.002>
66. Castaldo, L., Narváez, A., Izzo, L., Graziani, G., Gaspari, A., Di Minno, G., Ritieni, A.: Red Wine Consumption and Cardiovascular Health. *Molecules*. 24, 3626 (2019). <https://doi.org/10.3390/molecules24193626>
67. Gharakhanlou, B.J., Bonab, S.B., Amaghani, A., Shiri-Shahsavari, M.R.: Curcumin supplementation combined with high intensity interval training modulates serum irisin and lipid profile in obese women: “A randomized double-blind clinical trial.” *Contemp Clin Trials Commun*. 44, 101464 (2025). <https://doi.org/10.1016/j.conctc.2025.101464>
68. Zhang, W., Zheng, Y., Yan, F., Dong, M., Ren, Y.: Research progress of quercetin in cardiovascular disease. *Front Cardiovasc Med*. 10, 1203713 (2023). <https://doi.org/10.3389/fcvm.2023.1203713>
69. Han, D.-G., Seo, S.-W., Choi, E., Kim, M.-S., Yoo, J.-W., Jung, Y., Yoon, I.-S.: Impact of route-dependent phase-II gut metabolism and enterohepatic circulation on the bioavailability and systemic disposition of resveratrol in rats and humans: A comprehensive whole body physiologically-based pharmacokinetic modeling. *Biomedicine & Pharmacotherapy*. 151, 113141 (2022). <https://doi.org/10.1016/j.biopha.2022.113141>

70. Alghamdi, M.A., Fallica, A.N., Virzì, N., Kesharwani, P., Pittalà, V., Greish, K.: The Promise of Nanotechnology in Personalized Medicine. *J Pers Med.* 12, 673 (2022). <https://doi.org/10.3390/jpm12050673>
71. Awlqadr, F.H., Majeed, K.R., Altemimi, A.B., Hassan, A.M., Qadir, S.A., Saeed, M.N., Faraj, A.M., Salih, T.H., Abd Al-Manhel, A.J., Najm, M.A.A., Tsakali, E., Van Impe, J.F.M., Abd El-Maksoud, A.A., Abedelmaksoud, T.G.: Nanotechnology-based herbal medicine: Preparation, synthesis, and applications in food and medicine. *Journal of Agriculture and Food Research.* 19, 101661 (2025). <https://doi.org/10.1016/j.jafr.2025.101661>
72. Nwuruku, O.A., Ugwu, O.P.-C., Uti, D.E., Edwin, N.: Harnessing nature: plant-derived nanocarriers for targeted drug delivery in cancer therapy. *Phytomedicine Plus.* 5, 100828 (2025). <https://doi.org/10.1016/j.phyplu.2025.100828>
73. Atangwho, I.J., Egbung, G.E., Aja, P.M.: Lipid-based nano-carriers for the delivery of anti-obesity natural compounds: advances in targeted delivery and precision therapeutics. *Journal of Nanobiotechnology.* 23, 336 (2025). <https://doi.org/10.1186/s12951-025-03412-z>
74. Uti, D.E., Atangwho, I.J., Alum, E.U., Ntaobeten, E., Obeten, U.N., Bawa, I., Agada, S.A., Ukam, C.I.-O., Egbung, G.E.: Antioxidants in cancer therapy mitigating lipid peroxidation without compromising treatment through nanotechnology. *Discover Nano.* 20, 70 (2025). <https://doi.org/10.1186/s11671-025-04248-0>
75. Abbasi, H., Kouchak, M., Mirveis, Z., Hajipour, F., Khodarahmi, M., Rahbar, N., Handali, S.: What We Need to Know about Liposomes as Drug Nanocarriers: An Updated Review. *Adv Pharm Bull.* 13, 7–23 (2023). <https://doi.org/10.34172/apb.2023.009>
76. Lombardo, D., Kiselev, M.A.: Methods of Liposomes Preparation: Formation and Control Factors of Versatile Nanocarriers for Biomedical and Nanomedicine Application. *Pharmaceutics.* 14, 543 (2022). <https://doi.org/10.3390/pharmaceutics14030543>
77. Rao, L., Zhu, P., Guo, M., Hu, M., Guo, X., Du, Y., Xu, G.: Nebulized inhalation of nintedanib-loaded biomimetic nano-liposomes attenuated bleomycin-induced interstitial lung fibrosis in mice. *Nano Today.* 56, 102298 (2024). <https://doi.org/10.1016/j.nantod.2024.102298>
78. Uti, D.E., Egba, S.I., Ugwu, O.P.-C., Aja, P.M.: The Role of Phytochemicals in Age-Related Cognitive Decline: A Natural Solution for Brain Health. *Natural Product Communications.* 20, 1934578X251350761 (2025). <https://doi.org/10.1177/1934578X251350761>
79. Pandolfi, L., Marengo, A., Japiassu, K.B., Frangipane, V., Tsapis, N., Bincoletto, V., Codullo, V., Bozzini, S., Morosini, M., Lettieri, S., Vertui, V., Piloni, D., Arpicco, S., Fattal, E., Meloni, F.: Liposomes Loaded with Everolimus and Coated with Hyaluronic Acid: A Promising Approach for Lung Fibrosis. *Int J Mol Sci.* 22, 7743 (2021). <https://doi.org/10.3390/ijms22147743>
80. Ciupei, D., Colișar, A., Leopold, L., Stănilă, A., Diaconeasa, Z.M.: Polyphenols: From Classification to Therapeutic Potential and Bioavailability. *Foods.* 13, 4131 (2024). <https://doi.org/10.3390/foods13244131>
81. Capasso, L., De Masi, L., Sirignano, C., Maresca, V., Basile, A., Nebbioso, A., Rigano, D., Bontempo, P.: Epigallocatechin Gallate (EGCG): Pharmacological Properties, Biological Activities and Therapeutic Potential. *Molecules.* 30, 654 (2025). <https://doi.org/10.3390/molecules30030654>
82. Quesada-Vázquez, S., Eseberri, I., Les, F., Pérez-Matute, P., Herranz-López, M., Atgié, C., Lopez-Yus, M., Aranaz, P., Oteo, J.A., Escoté, X., Lorente-Cebrian, S., Roche, E., Courtois, A., López, V., Portillo, M.P., Milagro, F.I., Carpéné, C.: Polyphenols and metabolism: from present knowledge to future challenges. *J Physiol Biochem.* 80, 603–625 (2024). <https://doi.org/10.1007/s13105-024-01046-7>
83. Perrone, P., D'Angelo, S.: Gut Microbiota Modulation Through Mediterranean Diet Foods: Implications for Human Health. *Nutrients.* 17, 948 (2025). <https://doi.org/10.3390/nu17060948>
84. Jie, S., Fu, A., Wang, C., Rajabi, S.: A comprehensive review on the impact of polyphenol supplementation and exercise on depression and brain function parameters. *Behav Brain Funct.* 21, 10 (2025). <https://doi.org/10.1186/s12993-025-00273-2>
85. Orji OU, Ibiam UA, Aja PM, Ugwu P, Uraku AJ, Aloke C, Obasi OD, Nwali BU. Evaluation of the phytochemical and nutritional profiles of *Cnidioscolus aconitifolius* leaf collected in Abakaliki South East Nigeria. *World J Med Sci.* 2016;13(3):213-217.
86. Enechi OC, Okpe CC, Ibe GN, Omeje KO, Ugwu Okechukwu PC. Effect of *Buchholzia coriacea* methanol extract on haematological indices and liver function parameters in *Plasmodium berghei*-infected mice. *Glob Veterinaria.* 2016;16(1):57-66.
87. Alum EU, Uti DE, Ugwu Okechukwu PC, Alum BN. Toward a cure—Advancing HIV/AIDS treatment modalities beyond antiretroviral therapy: A review. *Med.* 2024;103(27):e38768.
88. Obeagu EI, Bot YS, Obeagu GU, Alum EU, Ugwu Okechukwu PC. Anaemia and risk factors in lactating mothers: A concern in Africa. *Int J Innov Appl Res.* 2024;11(2):15-17.

89. Alum EU, Ibiam UA, Ugwuja EI, Aja PM, Igwenyi IO, Offor CE, Orji UO, Ezeani NN, Ugwu OP, Aloke C, Egwu CO. Antioxidant effect of *Buchholzia coriacea* ethanol leaf extract and fractions on Freund's adjuvant-induced arthritis in albino rats: A comparative study. 2022;59(1):31-45.
90. Offor CE, Ugwu Okechukwu PC, Alum EU. Determination of ascorbic acid contents of fruits and vegetables. Int J Pharm Med Sci. 2015;5:1-3.
91. Amusa MO, Adepoju AO, Ugwu Okechukwu PC, Alum EU, Obeagu EI, Okon MB, Aja PM, Samson AOS. Effect of ethanol leaf extract of *Chromolaena odorata* on lipid profile of streptozotocin-induced diabetic Wistar albino rats. IAA J Biol Sci. 2024;10(1):109-117.
92. Enechi YS, Ugwu OC, Ugwu Okechukwu PC, Omeh K. Evaluation of the antinutrient levels of *Ceiba pentandra* leaves. IJRRPAS. 2013;3(3):394-400.
93. Ugwu Okechukwu PC, Nwodo OFC, Joshua EP, Odo CE, Ossai EC. Effect of ethanol leaf extract of *Moringa oleifera* on lipid profile of malaria-infected mice. Res J Pharm Biol Chem Sci. 2014;4(1):1324-1332.
94. OPC, Alum EU, Uhama KC. Dual burden of diabetes mellitus and malaria: Exploring the role of phytochemicals and vitamins in disease management. Res Inven J Res Med Sci. 2024;3(2):38-49.
95. Alum EU, Ugwu Okechukwu PC, Aja PM, Obeagu EI, Inya JE, Onyeije AP, Agu E, Awuchi CG. Restorative effects of ethanolic leaf extract of *Datura stramonium* against methotrexate-induced hematological impairments. Cogent Food Agric. 2013;9(1):2258774.
96. Offor CE, Nwankwegu FC, Joshua EP, Ugwu Okechukwu PC. Acute toxicity investigation and anti-diarrhoeal effect of the chloroform-methanol extract of the leaves of *Persea americana*. Iran J Pharm Res. 2014;13(2):651-658. PMID: 25237361; PMCID: PMC4157041.
97. Afiukwa CA, Oko AO, Afiukwa JN, Ugwu Okechukwu PC, Ali FU, Ossai EC. Proximate and mineral element compositions of five edible wild grown mushroom species in Abakaliki, southeast Nigeria. Res J Pharm Biol Chem Sci. 2013;4:1056-1064. Ugwu OP, Alum EU, Ugwu JN, Eze VH, Ugwu CN, Ogenyi FC, Okon MB. Harnessing technology for infectious disease response in conflict zones: Challenges, innovations, and policy implications. Med. 2024;103(28):e38834.
98. Obeagu EI, Ugwu OPC, Alum EU. Poor glycaemic control among diabetic patients; A review on associated factors. Newport Int J Res Med Sci (NIJRMS). 2023;3(1):30-33.
99. Nwaka AC, Ikechi-Agba MC, Okechukwu PU, Igwenyi IO, Agbafor KN, Orji OU, Ezugwu AL. The effects of ethanol extracts of *Jatropha curcas* on some hematological parameters of chloroform intoxicated rats. Am-Eur J Sci Res. 2015;10(1):45-49.
100. Ezeani NN, Ibiam UA, Orji OU, Igwenyi IO, Aloke C, Alum E, Aja PM, Ugwu OP. Effects of aqueous and ethanol root extracts of *Olax subscopioidea* on inflammatory parameters in complete Freund's adjuvant-collagen type II induced arthritic albino rats. Pharmacogn J. 2019;11(1)

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