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Dietary and Endogenous Antioxidants in Immune Homeostasis: A Translational Perspective

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ABSTRACT

Immune homeostasis relies on a dynamic equilibrium between oxidative stress, redox signaling, and the antioxidant systems that buffer reactive oxygen and nitrogen species generated during normal metabolism and inflammatory responses. Both endogenous antioxidants-such as glutathione, thioredoxin, uric acid, and enzymatic systems-and dietary antioxidants derived from fruits, vegetables, spices, and phytochemicals collectively maintain this balance. Disruption of redox homeostasis leads to immune dysfunction characterized by exaggerated inflammation, impaired pathogen clearance, altered lymphocyte differentiation, and progression of chronic metabolic and inflammatory diseases. This review synthesizes current mechanistic insights and emerging translational evidence on how dietary and endogenous antioxidants regulate innate and adaptive immunity through redox-sensitive pathways, including Nrf2, NF- κ B, HIF-1 α , and mitochondrial signaling. It further highlights clinical implications, therapeutic opportunities, and remaining research gaps required to bridge experimental knowledge with real-world clinical application. Understanding the interplay between antioxidant systems and immune regulation offers an evidence-based foundation for developing targeted dietary, pharmacological, and lifestyle interventions to restore immune balance in diverse pathological contexts.

Keywords: Antioxidants, Redox Homeostasis, Immune Regulation, Dietary Phytochemicals, Oxidative Stress

INTRODUCTION

Oxidative stress and immune function are deeply intertwined, forming a continuous feedback loop that influences nearly every phase of immune activity [1-5]. During immune activation, phagocytes and other leukocytes generate reactive oxygen species (ROS) and reactive nitrogen species (RNS) as essential antimicrobial agents and as intracellular signaling molecules that regulate gene expression, cytokine production, and cell differentiation [6-10]. Although these oxidants play indispensable physiological roles, their excess can rapidly shift the immune environment toward pathology. Elevated ROS levels promote lipid peroxidation, protein carbonylation, DNA strand breaks, mitochondrial dysfunction, and dysregulated inflammatory signaling [11-16]. These changes compromise cellular integrity and lead to chronic activation of inflammatory pathways, ultimately disrupting the delicate balance required for effective immunity [17-23]. To counter these potentially damaging effects, the body relies on a sophisticated array of endogenous antioxidant systems and complementary dietary antioxidants. Immune homeostasis, defined as the coordinated equilibrium between immune activation, tolerance, and resolution of inflammation, depends on the efficiency and responsiveness of these antioxidant defenses [24-29]. When antioxidant capacity is insufficient or overwhelmed-due to infection, metabolic stress, nutrient deficiencies, aging, or environmental exposures-redox imbalance drives the onset and progression of a wide spectrum of diseases [30-36]. These include autoimmune disorders, persistent infections, cardiometabolic diseases, certain cancers, and neuroinflammatory conditions [37-44]. Conversely, enhancing antioxidant status through endogenous

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upregulation or dietary intake can modulate immune signaling pathways, improve host resilience to pathogens, reduce tissue damage, and restore homeostatic balance [45-54].

A growing body of evidence demonstrates that redox regulation is not simply a protective mechanism against oxidative injury but also a fundamental determinant of immune cell development, differentiation, metabolism, and effector function [55-60]. Understanding how endogenous and dietary antioxidants interact with immune pathways provides a powerful framework for developing nutritional and pharmacological interventions that promote health and prevent chronic disease [61-67]. This review integrates mechanistic, nutritional, and translational perspectives to highlight the roles of endogenous antioxidant systems in immune regulation and their broader implications for clinical practice and public health.

2. Endogenous Antioxidant Systems in Immune Regulation

2.1 Glutathione System

Glutathione (GSH) is the most abundant intracellular antioxidant and a central regulator of redox-sensitive immune processes [68-74]. The reduced form of glutathione directly neutralizes ROS and supports the activity of glutathione peroxidases, which detoxify hydrogen peroxide and lipid peroxides. The ratio of reduced to oxidized glutathione is a widely used indicator of cellular redox balance and immune competence [75-80]. GSH controls T cell activation by regulating redox-sensitive transcription factors, supports antigen presentation by protecting dendritic cells from oxidative injury, influences macrophage polarization toward the anti-inflammatory M2 pathway, and safeguards lymphocytes from activation-induced cell death. Deficiency in GSH impairs lymphocyte proliferation, reduces cytokine production, and increases susceptibility to infections.

2.2 Thioredoxin and Peroxiredoxin Systems

Thioredoxin and peroxiredoxins form a complementary antioxidant network that maintains the redox state of protein thiols [81-84]. These systems play key roles in DNA synthesis during lymphocyte expansion, regulation of inflammasome activation, and modulation of cytokine signaling via reversible oxidation of cysteine residues [85-90]. Extracellular thioredoxin also functions as a cytokine-like molecule capable of influencing leukocyte recruitment and chemotaxis, linking redox buffering to immune communication.

2.3 Enzymatic Antioxidants

Superoxide dismutase, catalase, and glutathione peroxidases constitute the core enzymatic antioxidant machinery [91-96]. Together, they detoxify superoxide radicals and hydrogen peroxide, protect immune cells from self-inflicted oxidative damage during respiratory bursts, and preserve the integrity of epithelial and endothelial barriers essential for innate defense [91-96].

2.4 Endogenous Small-Molecule Antioxidants

Small molecules such as uric acid, bilirubin, lipoic acid, and coenzyme Q10 contribute significantly to systemic antioxidant capacity [17]. These molecules possess strong radical-scavenging abilities and modulate inflammation, mitochondrial function, and vascular homeostasis [18]. Despite their modest size, their physiological influence on redox and immune regulation is substantial, supporting the broader antioxidant network that sustains immune homeostasis.

3. Dietary Antioxidants and Their Impact on Immunity

Dietary antioxidants comprise a wide range of nutrients and bioactive compounds, including vitamins such as C, E, and A, trace minerals like selenium and zinc, polyphenols, carotenoids, and various plant-derived phytochemicals [19]. Together, these molecules fortify endogenous antioxidant systems and exert direct regulatory effects on immune pathways [20]. Their influence extends beyond simple free radical scavenging; many act as signaling modulators that shape cellular differentiation, cytokine release, membrane stability, and gene expression patterns involved in both innate and adaptive immunity [21]. Adequate intake of these antioxidants is therefore essential for maintaining immune resilience, preventing chronic inflammation, and supporting recovery from infection or tissue injury.

3.1 Vitamin C (Ascorbate)

Vitamin C is one of the most important water-soluble antioxidants and accumulates in leukocytes at concentrations far higher than in plasma [22]. It enhances innate immunity by improving neutrophil chemotaxis, promoting the formation of reactive oxygen species required for microbial killing, and facilitating clearance of spent neutrophils from inflammatory sites [23]. Vitamin C also regenerates oxidized vitamin E and sustains glutathione levels, amplifying systemic antioxidant capacity. By modulating cytokine production and dampening excessive inflammatory responses, it contributes to balanced immunity. Deficiency leads to impaired epithelial barrier function, reduced lymphocyte activation, and an increased susceptibility to infections.

3.2 Vitamin E (Tocopherols and Tocotrienols)

Vitamin E is a lipid-soluble antioxidant that protects cellular membranes from peroxidation [24]. It improves T cell responsiveness by maintaining membrane fluidity and enhancing signal transduction. Vitamin E supplementation has been shown to reduce age-related immune decline, partly by revitalizing T cell-mediated responses [25]. Its regulatory effects on prostaglandin synthesis and inflammatory signaling support a more controlled inflammatory response and improved immune coordination.

3.3 Vitamin A and Carotenoids

Vitamin A and its derivatives play central roles in mucosal immunity, promoting epithelial integrity, supporting IgA secretion, and guiding the differentiation of lymphocyte subsets [26]. Retinoids influence the balance between Th1, Th2, and regulatory T cells, thereby shaping adaptive immunity [27]. Carotenoids such as beta-carotene enhance natural killer cell activity, improve antioxidant capacity within immune cells, and contribute to resistance against oxidative stress encountered during immune activation.

3.4 Selenium

Selenium is indispensable for the activity of glutathione peroxidases and thioredoxin reductases, enzymes crucial for maintaining cellular redox stability [28]. Selenium deficiency compromises lymphocyte proliferation, weakens antiviral defenses, and heightens vulnerability to oxidative damage during inflammation [29]. Optimal selenium intake enhances cytotoxic T cell and NK cell activity, supporting both innate and adaptive immune responses.

3.5 Polyphenols

Polyphenols, widely found in fruits, vegetables, teas, and spices, are among the most potent dietary modulators of immune function [30]. Compounds such as quercetin, catechins, curcumin, and resveratrol operate through multiple mechanisms. They neutralize ROS, activate the Nrf2 pathway to induce endogenous antioxidant genes, inhibit NF-κB-driven inflammatory cytokine expression, and promote anti-inflammatory macrophage phenotypes [31]. Many polyphenols also facilitate the development of regulatory T cells, enhancing immune tolerance and helping to prevent chronic inflammatory disorders.

3.6 Dietary Fiber and Microbiota-Derived Antioxidants

Dietary fiber indirectly contributes to antioxidant and immune regulation through fermentation by gut microbiota [32]. The resulting short-chain fatty acids, especially butyrate, serve as metabolic substrates for intestinal cells, strengthen epithelial barrier function, modulate inflammatory gene expression, and act as epigenetic regulators [33]. Through these mechanisms, fiber-derived metabolites enhance systemic antioxidant defenses and sustain optimal immune homeostasis.

5. Antioxidant Regulation of Innate Immunity

Innate immune cells are particularly sensitive to redox conditions. In macrophages, oxidative stress favors pro-inflammatory M1 polarization, whereas reducing environments promote M2 phenotypes involved in tissue repair and resolution [34]. Dietary antioxidants therefore support balanced macrophage responses.

Neutrophils rely on ROS for antimicrobial activity, yet excessive generation damages host tissues. Antioxidants improve the efficiency of microbial killing, protect neutrophils from premature apoptosis, and limit collateral tissue injury.

Natural killer cells require an optimal redox environment for cytotoxicity and cytokine release. Nutrients such as vitamins C and E, as well as polyphenols like quercetin and resveratrol, enhance NK cell performance.

Dendritic cells also depend on controlled redox signaling for antigen processing and T cell priming. Elevated ROS impair their function, while antioxidants help maintain antigen presentation and promote regulatory immune responses.

6. Antioxidant Regulation of Adaptive Immunity

6.1 T Lymphocytes

T lymphocyte function is tightly coupled to intracellular redox status, as activation, differentiation, and survival all depend on controlled ROS signaling. Low to moderate ROS levels act as second messengers that facilitate TCR signaling and metabolic reprogramming, but excessive ROS disrupt these processes [35]. High oxidative burden inhibits proliferation by blocking key kinase cascades, reduces IL-2 and other cytokines, and can drive T cells toward anergy or apoptosis through mitochondrial damage. Adequate antioxidant buffering preserves the signaling window necessary for effective T cell responses. Dietary and endogenous antioxidants promote balanced Th1/Th2 differentiation, support the expansion and suppressive function of regulatory T cells, and counteract the differentiation of pro-inflammatory Th17 cells [36]. Redox homeostasis also enhances the formation and longevity of memory T cells by sustaining mitochondrial fitness and preventing oxidative exhaustion, improving long-term immune protection.

6.2 B Lymphocytes

B cell maturation, germinal-center reactions, and antibody class switching rely on precise redox control. Antioxidants help maintain the integrity of B cell receptor signaling and support the survival of activated B cells [37]. Polyphenols and vitamins A and E are particularly influential, as they enhance B cell responsiveness, improve somatic hypermutation efficiency, and promote the generation of high-affinity antibodies. By stabilizing cellular redox balance, antioxidants ultimately reinforce humoral immunity and contribute to more robust, durable antibody-mediated protection.

CONCLUSION

Antioxidants—both endogenous and dietary—play indispensable roles in maintaining immune homeostasis through regulation of redox-sensitive pathways, modulation of immune cell function, and preservation of tissue integrity. Their influence spans innate and adaptive immunity, metabolic and inflammatory signaling, and resilience against infections and chronic disease. Translational evidence supports targeted use of antioxidants as complementary strategies to enhance immune function, reduce inflammation, and improve health outcomes. Continued integrative research bridging molecular biology, nutrition, and clinical medicine will advance the development of precision antioxidant therapies tailored to individual redox and immunological profiles.

REFERENCES

1. Alum, E.U., Uti, D.E. & Ofor, C.E. Redox Signaling Disruption and Antioxidants in Toxicology: From Precision Therapy to Potential Hazards. *Cell Biochem Biophys* (2025). <https://doi.org/10.1007/s12013-025-01846-8>
2. Dery KJ, Chiu R, Kasargod A, Kupiec-Weglinski JW. Feedback Loops Shape Oxidative and Immune Interactions in Hepatic Ischemia–Reperfusion Injury. *Antioxidants*. 2025; 14(8):944. <https://doi.org/10.3390/antiox14080944>
3. Ochulor Okechukwu C., Njoku Obioma U., Uroko Robert I and Egba Simeon I. Nutritional composition of *Jatropha tanjorensis* leaves and effects of its aqueous extract on carbon tetrachloride induced oxidative stress in male Wistar albino rats. *Biomedical Research* 2018; 29(19): 3569-3576
4. Iddir M, Brito A, Dingo G, Fernandez Del Campo SS, Samouda H, La Frano MR, Bohn T. Strengthening the Immune System and Reducing Inflammation and Oxidative Stress through Diet and Nutrition: Considerations during the COVID-19 Crisis. *Nutrients*. 2020; 12(6):1562. <https://doi.org/10.3390/nu12061562>
5. Ugwu, CE., Sure, SM., Dike, CC., Okpoga, NA and Egba, SI. Phytochemical and *in vitro* antioxidant activities of methanol leave extract of *Alternanthera basiliana*. *Journal of Pharmacy Research*, 2018; 12(6): 835-839
6. Ogugua, Victor N., Njoku, Obioma U., Egba, Simeon I., Uroko, Robert I and Ignatius Glory. In vitro study of nutritional and antioxidant properties of methanol extract of *Nauclea latifolia* root bark. *Biomedical Research*, 2018; 29(21): 3766-3773
7. Alum, E.U., Nwuruku, A.O. and Edwin, N. Targeting Oxidative Stress in Cancer Management: The Role of Antioxidant Phytochemicals. *KIU J. Health Sci.*, 4(2): 1-10. <https://doi.org/10.59568/KJHS-2024-4-2-01>
8. Silvestrini A, Mancini A. The Double-Edged Sword of Total Antioxidant Capacity: Clinical Significance and Personal Experience. *Antioxidants*. 2024; 13(8):933. <https://doi.org/10.3390/antiox13080933>
9. Bellanti F, Coda ARD, Trecca MI, Lo Buglio A, Serviddio G, Vendemiale G. Redox Imbalance in Inflammation: The Interplay of Oxidative and Reductive Stress. *Antioxidants (Basel)*. 2025 May 29;14(6):656. doi: 10.3390/antiox14060656. PMID: 40563291; PMCID: PMC12189482.
10. Uhwo E N, Egba S I, Nwuke P C, Obike C A and Kelechi G K. Antioxidative properties of *Adansonia digitata* L. (baobab) leaf extract exert protective effect on doxorubicin induced cardiac toxicity in Wistar rats. *Clinical Nutrition Open Science* 2022; 45:3-16
11. Aoyama K, Nakaki T. Glutathione in Cellular Redox Homeostasis: Association with the Excitatory Amino Acid Carrier 1 (EAAC1). *Molecules*. 2015 May 14;20(5):8742-58. doi: 10.3390/molecules20058742. PMID: 26007177; PMCID: PMC6272787.
12. Matuz-Mares D, Riveros-Rosas H, Vilchis-Landeros MM, Vázquez-Meza H. Glutathione Participation in the Prevention of Cardiovascular Diseases. *Antioxidants*. 2021; 10(8):1220. <https://doi.org/10.3390/antiox10081220>
13. Netto LE, Antunes F. The Roles of Peroxiredoxin and Thioredoxin in Hydrogen Peroxide Sensing and in Signal Transduction. *Mol Cells*. 2016 Jan;39(1):65-71. doi: 10.14348/molcells.2016.2349. Epub 2016 Jan

25. PMID: 26813662; PMCID: PMC4749877.
14. Oberacker T, Kraft L, Schanz M, Latus J, Schrickler S. The Importance of Thioredoxin-1 in Health and Disease. *Antioxidants*. 2023; 12(5):1078. <https://doi.org/10.3390/antiox12051078>
15. Jomova K, Alomar SY, Alwasel SH, Nepovimova E, Kuca K, Valko M. Several lines of antioxidant defense against oxidative stress: antioxidant enzymes, nanomaterials with multiple enzyme-mimicking activities, and low-molecular-weight antioxidants. *Arch Toxicol*. 2024 May;98(5):1323-1367. doi: 10.1007/s00204-024-03696-4. Epub 2024 Mar 14. PMID: 38483584; PMCID: PMC11303474.
16. Ibiam, U. A., Ugwuja, E. I., Aja, P. M., Igwenyi, I. O., Offor, C. E., Orji, O. U., Ezeani N. N, Ugwu, O. P. C., Aloke, C., Egwu, C. O. Antioxidant Effect of *Buchholzia coriacea* Ethanol Leaf Extract and Fractions on Freund's Adjuvant-induced Arthritis in Albino Rats: A Comparative Study. *Slovenian Veterinary Research*. 2022; 59 (1): 31–45. doi: 10.26873/svr-1150-2022.
17. Silvestri S, Orlando P, Armeni T, Padella L, Bruguè F, Seddaiu G, Littarru GP, Tiano L. Coenzyme Q10 and α -lipoic acid: antioxidant and pro-oxidant effects in plasma and peripheral blood lymphocytes of supplemented subjects. *J Clin Biochem Nutr*. 2015 Jul;57(1):21-6. doi: 10.3164/jcbn.14-130. Epub 2015 Apr 16. PMID: 26236096; PMCID: PMC4512890.
18. Ogbonna OA., Egba, SI., Uhwo EN., Omeoga HC., Obeagu EI. Toxic outcomes of ciprofloxacin and gentamicin co-administration and possible ameliorating role for antioxidant vitamins C and E in Wistar Rats. *Elite Journal of Medicine*, 2024; 2(3): 1-14.
19. Gulcin İ. Antioxidants: a comprehensive review. *Arch Toxicol*. 2025 May;99(5):1893-1997. doi: 10.1007/s00204-025-03997-2. Epub 2025 Apr 15. PMID: 40232392; PMCID: PMC12085410.
20. Manful CF, Fordjour E, Subramaniam D, Sey AA, Abbey L, Thomas R. Antioxidants and Reactive Oxygen Species: Shaping Human Health and Disease Outcomes. *International Journal of Molecular Sciences*. 2025; 26(15):7520. <https://doi.org/10.3390/ijms26157520>
21. Andrés CMC, Pérez de la Lastra JM, Juan CA, Plou FJ, Pérez-Lebeña E. Antioxidant Metabolism Pathways in Vitamins, Polyphenols, and Selenium: Parallels and Divergences. *Int J Mol Sci*. 2024 Feb 23;25(5):2600. doi: 10.3390/ijms25052600. PMID: 38473850; PMCID: PMC10932048.
22. Alberts A, Moldoveanu E-T, Niculescu A-G, Grumezescu AM. Vitamin C: A Comprehensive Review of Its Role in Health, Disease Prevention, and Therapeutic Potential. *Molecules*. 2025; 30(3):748. <https://doi.org/10.3390/molecules30030748>
23. Aja, W., Ugwu, O. P. C., Obeagu, E. I., Okon, M. B. Assessment of vitamin composition of ethanol leaf and seed extracts of *Datura stramonium*. *Avicenna J Med Biochem*. 2023; 11(1):92-97. doi:10.34172/ajmb.2023.2421.
24. Rizvi S, Raza ST, Ahmed F, Ahmad A, Abbas S, Mahdi F. The role of vitamin e in human health and some diseases. *Sultan Qaboos Univ Med J*. 2014 May;14(2):e157-65. Epub 2014 Apr 7. PMID: 24790736; PMCID: PMC3997530.
25. Kilcarslan You D, Fuwad A, Lee KH, Kim HK, Kang L, Kim SM, Jeon T-J. Evaluation of the Protective Role of Vitamin E against ROS-Driven Lipid Oxidation in Model Cell Membranes. *Antioxidants*. 2024; 13(9):1135. <https://doi.org/10.3390/antiox13091135>
26. Huang Z, Liu Y, Qi G, Brand D, Zheng SG. Role of Vitamin A in the Immune System. *J Clin Med*. 2018 Sep 6;7(9):258. doi: 10.3390/jcm7090258. PMID: 30200565; PMCID: PMC6162863.
27. De Medeiros PHQS, Pinto DV, De Almeida JZ, Rêgo JMC, Rodrigues FAP, Lima AÂM, Bolick DT, Guerrant RL, Oriá RB. Modulation of Intestinal Immune and Barrier Functions by Vitamin A: Implications for Current Understanding of Malnutrition and Enteric Infections in Children. *Nutrients*. 2018; 10(9):1128. <https://doi.org/10.3390/nu10091128>
28. Lee JG, Jang JY, Baik SM. Selenium as an Antioxidant: Roles and Clinical Applications in Critically Ill and Trauma Patients: A Narrative Review. *Antioxidants (Basel)*. 2025 Feb 28;14(3):294. doi: 10.3390/antiox14030294. PMID: 40227249; PMCID: PMC11939285.
29. Shahidin, Wang Y, Wu Y, Chen T, Wu X, Yuan W, Zhu Q, Wang X, Zi C. Selenium and Selenoproteins: Mechanisms, Health Functions, and Emerging Applications. *Molecules*. 2025; 30(3):437. <https://doi.org/10.3390/molecules30030437>
30. Alum, E.U. Unlocking the Secrets of Nature: Phytochemicals as Key Players in Longevity and Healthy Aging. *Cell Biochem Biophys* (2025). <https://doi.org/10.1007/s12013-025-01872-6>

31. Akwari, A.A., Okoroh, P.N., Aniokete, U.C., Abba, J.N., Uti, D.E. Phytochemicals as modulators of ferroptosis: a novel therapeutic avenue in cancer and neurodegeneration. *Mol Biol Rep* **52**, 636 (2025). <https://doi.org/10.1007/s11033-025-10752-4>
32. Aniokete, U. C., Emeruwa, A. P., Obasi, D. C., Okoroh, P. N., ... & Aja, P. M. (2025). Gut microbiota at the crossroads of food additives, pollutants, and chronic disease risk. *Toxicology and Environmental Health Sciences*. <https://doi.org/10.1007/s13530-025-00295-3>
33. Ugwu, O.P., Okon, M.B., Alum, E.U., Ugwu, C.N., Anyanwu, E.G., Mariam, B., et al. (2025). Unveiling the therapeutic potential of the gut microbiota-brain axis: Novel insights and clinical applications in neurological disorders. *Medicine (Baltimore)*. 2025 Jul 25;104(30):e43542. doi: 10.1097/MD.00000000000043542. PMID: 40725913; PMCID: PMC12303509.
34. Pérez S, Rius-Pérez S. Macrophage Polarization and Reprogramming in Acute Inflammation: A Redox Perspective. *Antioxidants (Basel)*. 2022 Jul 19;11(7):1394. doi: 10.3390/antiox11071394. PMID: 35883885; PMCID: PMC9311967.
35. Peng HY, Lucavs J, Ballard D, Das JK, Kumar A, Wang L, Ren Y, Xiong X, Song J. Metabolic Reprogramming and Reactive Oxygen Species in T Cell Immunity. *Front Immunol*. 2021 Mar 31;12:652687. doi: 10.3389/fimmu.2021.652687. PMID: 33868291; PMCID: PMC8044852.
36. Lindsay RT, Rhodes CJ. Reactive Oxygen Species (ROS) in Metabolic Disease—Don't Shoot the Metabolic Messenger. *International Journal of Molecular Sciences*. 2025; 26(6):2622. <https://doi.org/10.3390/ijms26062622>
37. Kurutas EB. The importance of antioxidants which play the role in cellular response against oxidative/nitrosative stress: current state. *Nutr J*. 2016 Jul 25;15(1):71. doi: 10.1186/s12937-016-0186-5. PMID: 27456681; PMCID: PMC4960740.
38. Ugwu OPC, Nwodo OFC, Joshua PE, Odo CE, Ossai EC, Abubakar B. Phytochemical and acute toxicity studies of Moringa oleifera ethanol leaf extract. *Int J Life Sci Biotechnol Pharma Res*. 2013;2(1).
39. Ugwu OPC, Nwodo OFC, Joshua PE, Odo CE, Ossai EC. The effect of ethanol leaf extract of Moringa oleifera on the lipid profile of malaria infected mice. *Res J Pharm Biol Chem Sci*. 2013;4(1).
40. Ugwu OPC, Nwodo OFC, Joshua PE, Odo CE, Ossai EC, Abubakar B. Ameliorative effects of ethanol leaf extract of Moringa oleifera on the liver and kidney markers of malaria infected mice. *Int J Life Sci Biotechnol Pharma Res*. 2013;2(1).
41. Ugwu OPC, Nwodo OFC, Joshua PE, Odo CE, Bawa A, Ossai EC. Anti-malaria and hematological analyses of ethanol leaf extract of Moringa oleifera on malaria infected mice. *Int J Pharm Biol Sci*. 2013;3(1):360–71.
42. Enechi OC, Manyawo LN, Ugwu OPC. Effect of ethanol seed extract of *Bucchozia coriacea* (wonderful kola) on the lipid profile of albino rats. *World J Pharm Pharm Sci*. 2013;2(3):802–13.
43. Enechi OC, Oluka IH, Ugwu OPC, Omeh YS. Effect of ethanol leaf extract of *Alstonia boonei* on the lipid profile of alloxan induced diabetic rats. *World J Pharm Pharm Sci*. 2013;2(3):782–95.
44. Enechi OC, Peter CD, Ugwu OPC, Udeh SMC, Omeh YS. Evaluation of the nutritional potential of *Ceiba pentandra* leaves. *Mintage J Pharm Med Sci*. 2013;2(3):25–7.
45. Enechi OC, Igbonekwu CN, Ugwu OPC. Effects of ethanol extract of *Cissus quadrangularis* on induced gastric ulcer in rats. *Afr J Biotechnol*. 2013;12(43):6197–202.
46. Enechi OC, Obiora EN, Ugwu OPC. Chromatographic identification and the effect of the alkaloidal extract of *Bucchozia coriacea* seeds on the body weights and relative liver weights of mice. *Adv Biol Res*. 2013;7(5):188–93.
47. Afiukwa CA, Ugwu OPC, Ebenyi LN, Ossai EC, Nwaka AC. Phytochemical analysis of three wild edible mushrooms, coral mushroom, *Agaricus bisporus* and *Lentinus sajor caju*, common in Ohaukwu Area of Ebonyi State, Nigeria. *Int J Pharm*. 2013;3(2):410–4.

48. Afiukwa CA, Ugwu OPC, Okoli SO, Idenyi JN, Ossai EC. Contents of some vitamins in five edible mushroom varieties consumed in Abakaliki Metropolis, Nigeria. *Res J Pharm Biol Chem Sci.* 2013;4(2).
49. Afiukwa CA, Oko AO, Afiukwa JN, Ugwu OPC, Ali FU, Ossai EC. Proximate and mineral element compositions of five edible wild grown mushroom species in Abakaliki, southeast Nigeria. *Res J Pharm Biol Chem Sci.* 2013;4(2):1055.
50. Afiukwa CA, Ogah O, Ugwu OPC, Oguguo JO, Ali FU, Ossai EC. Nutritional and antinutritional characterization of two wild yam species from Abakaliki, southeast Nigeria. *Res J Pharm Biol Chem Sci.* 2013;4(2).
51. Adonu CC, Ugwu OPC, Esimone CO, Ossai EC, Bawa A, Nwaka AC. Phytochemical analyses of the methanol, hot water and N-hexane extracts of the aerial parts of *Cassytha filiformis* (Linn) and leaves of *Cleistopholis patens* (Benth). *Int J Pharm Biol Chem Sci.* 2013;3(1).
52. Adonu CC, Esimone CO, Ugwu OPC, Bawa A, Ossai EC. In vitro evaluation of the antibacterial potential of extracts of the aerial parts of *Cassytha filiformis* against urogenital clinical gram-positive organisms. *Int J Pharm Biol Chem Sci.* 2013;3(1).
53. Enechi OC, Ogochukwu BO, Ugwu OPC. Effect of fermentation on biochemical properties of maize (*Zea mays* L.). 2014.
54. Enechi OC, Stephen A, Ugwu OPC. Concentrations of iodine and some environmental goitrogens in two selected water bodies—Adada and Akoru in Nsukka, Enugu State, Nigeria. *Afr J Biotechnol.* 2014;13(44):4215–9.
55. Udeozo IP, Nwaka AC, Ugwu OPC, Akogwu M. Anti-inflammatory, phytochemical and acute toxicity study of the flower extract of *Newbouldia laevis*. *Int J Curr Microbiol App Sci.* 2014;3(3):1029–35.
56. Enechi OC, Oluka HI, Ugwu OPC. Acute toxicity and ameliorative properties of *Alstonia boonei* leaf extract on diabetic rats. *Afr J Biotechnol.* 2014;13(5).
57. Enechi OC, Oluka HI, Ugwu OPC. Acute toxicity, lipid peroxidation and ameliorative properties of *Alstonia boonei* ethanol leaf extract on the kidney markers of alloxan induced diabetic rats. *Afr J Biotechnol.* 2014;13(5).
58. Odo CE, Nwodo OFC, Joshua PE, Ugwu OPC, Okonkwo CC. Acute toxicity investigation and anti-diarrhoeal effect of the chloroform-methanol extract of the seeds of *Persea americana* in albino rats. *J Pharm Res.* 2013;6(3):331–5.
59. Odo CE, Nwodo OFC, Joshua PE, Ugwu OPC. Acute toxicity investigation and anti-diarrhoeal effect of the chloroform-methanol extract of the leaves of *Persea americana*. *Iran J Pharm Res.* 2014;13(2):651–8.
60. Amalu PC, Chukwuezi FO, Ugwu OPC. Antimicrobial effects of bitter kola (*Garcinia kola*) nut on *Staphylococcus aureus*, *Escherichia coli* and *Candida albicans*. *IOSR J Dent Med Sci.* 2014;13(4):29–32.
61. Ilozue NM, Ikezu UP, Ugwu OPC. Antimicrobial and phytochemical screening of the seed extracts of *Persea americana* (avocado pear). *IOSR J Pharm Biol Sci.* 2014;9(2):23–5.
62. Orji OU, Ibiam UA, Aja PM, Ugwu OPC, Uraku AJ, Inya-Agha OR, et al. Evaluation of the phytochemical and nutritional profiles of *Cnidioscolus aconitifolius* leaf collected in Abakaliki, south east Nigeria. *World J Med Sci.* 2015;13(3):213–7.
63. Igwenyi IO, Nchi PO, Ugwu OPC, Igwenyi IP, Obasi DC, Edwin N, et al. Nutritional potential of *Azadirachta indica* seeds. *Indo Am J Pharm Sci.* 2017;4(2):477–82.
64. Aja PM, Ugwu OPC, Kennedy K, Ibere JB, Ekpono EU. Phytochemical analysis of *Senna occidentalis* leaves. *IDOSR J Appl Sci.* 2017;2(1):75–91.

65. Ibiam UA, Alum EU, Orji OU, Aja PM, Ezeani NN, Ugwu OPC, et al. Anti-inflammatory effects of *Buchholzia coriacea* ethanol leaf-extract and fractions in Freund's adjuvant-induced rheumatoid arthritic albino rats. *Indo Am J Pharm Sci.* 2018;5(7):6341–57.
66. Ibiam UA, Alum EU, Aja PM, Orji OU, Ezeani NN, Ugwu OPC, et al. Comparative analysis of chemical composition of *Buchholzia coriacea* ethanol leaf-extract, aqueous and ethylacetate fractions. *Indo Am J Pharm Sci.* 2018;5(7):6358–69.
67. Ezeani NN, Ibiam UA, Orji OU, Igwenyi IO, Alope C, Alum E, et al. Effects of aqueous and ethanol root extracts of *Olox subscorpioidea* on inflammatory parameters in complete Freund's adjuvant-collagen type II induced arthritic albino rats. *Pharmacognosy J.* 2019;11(1).
68. Alum EU. Antioxidant effect of *Buchholzia coriacea* ethanol leaf-extract and fractions on Freund's adjuvant-induced arthritis in albino rats: a comparative study. *Slov Vet Res.* 2022;59(1).
69. Alum EU, Inya JE, Ugwu OPC, Obeagu EI, Alope C, Aja PM, et al. Ethanolic leaf extract of *Datura stramonium* attenuates methotrexate-induced biochemical alterations in Wistar albino rats. *RPS Pharm Pharmacol Rep.* 2023;2(1):1–6.
70. Alum EU, Ugwu OPC, Aja PM, Obeagu EI, Inya JE, Onyeije AP, et al. Restorative effects of ethanolic leaf extract of *Datura stramonium* against methotrexate-induced hematological impairments. *Cogent Food Agric.* 2023;9(1):2258774.
71. Alum EU, Oyika MT, Ugwu OPC, Aja PM, Okon MB, Diana MC, et al. Comparative analysis of mineral constituents of ethanol leaf and seed extracts of *Datura stramonium*. *IDOSR J Appl Sci.* 2023;8(1):143–51.
72. Alum EU, Diana MC, Ugwu OPC, Aja PM, Okon MB. Phytochemical composition of *Datura stramonium* ethanol leaf and seed extracts: a comparative study. *IAA J Biol Sci.* 2023;10(1):118–25.
73. Alum EU, Ugwu OPC, Obeagu EI, Aja PM, Okon MB. Assessment of vitamin composition of ethanol leaf and seed extracts of *Datura stramonium*. *Avicenna J Med Biochem.* 2023;11(1):92–7.
74. Alum EU, Manjula VS, Uti DE, Echegu DA, Ugwu OPC, Egba SI, et al. Metabolomics-driven standardization of herbal medicine: advances, applications, and sustainability considerations. *Nat Prod Commun.* 2025;20(8):1934578X251367650.
75. Alum EU, Obasi DC, Abba JN, Aniokete UC, Okoroh PN, Ugwu OPC, et al. Endogenous plant signals and human health: molecular mechanisms, ecological functions, and therapeutic prospects. *Biochem Biophys Rep.* 2025;43:102114.
76. Alum EU, Nwuruku OA, Ugwu OPC, Uti DE, Alum BN, Edwin N. Harnessing nature: plant-derived nanocarriers for targeted drug delivery in cancer therapy. *Phytomedicine Plus.* 2025;5(3):100828.
77. Alum EU, Uti DE, Ugwu OPC, Okon MB, Aggad WS, Basajja M, et al. Medicinal plants and the gastrointestinal microbiota in chronic diseases modulation: a structured mechanistic and translational review. *Curr Microbiol.* 2026;83(6):346.
78. Alum EU, Manjula VS, Ugwu OPC, Uti DE, Alum BN, Echegu DA. The Internet of Plants: re-imagining plant signalling networks through an information-and-communication theory lens. *Nat Prod Commun.* 2026;21(5):1934578X261455751.
79. Ugwu OPC, Ugwu MN, Onohuean H, Rather HA, Usman IM. Noncoding RNAs and the phytochemical economy: molecular regulators of secondary metabolism in medicinal plants. *Biochem Biophys Rep.* 2026;45:102486.
80. Ugwu OPC, Ugwu MN, Ogenyi FC, Eze VHU, Alum EU. Nano-enhanced phytomedicine: a review of nanocarrier systems for targeted delivery of plant-derived bioactives in chronic disease therapy. *Phytomedicine Plus.* 2026.

81. Alum EU, Nwuruku OA, Uti DE, Echegu DA, Ugwu OPC, Edwin N, et al. Unlocking the potential of endophytes in enhancing plant secondary metabolite biosynthesis. *Biochem Biophys Rep.* 2026;45:102385.
82. Ugwu OPC, Ogenyi FC, Alum EU, Basajja M, Ugwu CN, Ugwu JN, et al. Plant- and soil-derived polyphenols shape soil microbiomes and crop outcomes: a systematic review and meta-analysis. *Front Soil Sci.* 2026;6. doi:10.3389/fsoil.2026.17531
83. Mbyeire H, Fasogbon IV, Musyoka AM, Oviosun A, Ojiakor VO, Ugwu OPC, et al. Exploring the use of phytotherapy in benign prostatic hyperplasia [BPH]: a systematic review. *F1000Res.* 2026;14:412.
84. Ugwu OPC, Anyanwu CN, Ugwu MN. Harnessing plant metabolic pathways for innovative diabetes management: unlocking the therapeutic potential of medicinal plants. *Plant Signal Behav.* 2025;20(1):e2486076.
85. Uti DE, Atangwho IJ, Alum EU, Egba SI, Ugwu OPC, Ikechukwu GC, et al. Natural antidiabetic agents: current evidence and development pathways from medicinal plants to clinical use. *Nat Prod Commun.* 2025;20(3):1–29.
86. Ugwu OPC, Alum EU, Kungu E, Inyangat R, Obeagu EI. Exploring indigenous medicinal plants for managing diabetes mellitus in Uganda: ethnobotanical insights, pharmacotherapeutic strategies, and national development alignment. *INOSR Exp Sci.* 2023;12(2):214–24.
87. Ugwu OPC, Alum EU, Obeagu EI. Integrating medicinal plant diversity in post-COVID Uganda for holistic healthcare management. *IAA J Biol Sci.* 2023;10(3):32–41.
88. Ugwu OPC, Alum EU, Kungu E, Inyangat R. Exploration of medicinal plants used in the management of malaria in Uganda. *Newport Int J Res Med Sci.* 2023;4(1):101–8.
89. Alum EU, Ugwu OPC. Beyond nutrients: exploring the potential of phytochemicals for human health. *IAA J Appl Sci.* 2023;10(3):1–7.
90. Alum EU, Uti DE, Egba SI, Ugwu OPC, Aja PM. The role of phytochemicals in age-related cognitive decline: a natural solution for brain health. *Nat Prod Commun.* 2025;20(6):1–18.
91. Ogbodo JO, Egba SI, Ikechukwu GC, Paul PC, Mba JO, Ugwu OPC, et al. Volatile organic compound-drug receptor interactions: a potential tool for drug design in the search for remedies for increasing toxic occupational exposure. *Processes.* 2025;13(1):154.
92. Ikuomola EO, Owu DU, Oka VO, Aja PM, Ugwu NF, Umar US, et al. A review of medicinal plants used for the restoration of reproductive functionality following cimetidine-induced reproductive toxicity. *RPS Pharm Pharmacol Rep.* 2024;3(3):rqae014. doi:10.1093/rpsppr/rqae014
93. Ikuomola EO, Owu DU, Oka VO, Agba S, Aja PM. Gas chromatography-mass spectrometric (GC-MS) revealed bioactive constituents of *Brassica oleracea* var. *viridis* (collard greens) used as ethnomedicine to treat male infertility and related conditions. *F1000Res.* 2025;14:525.
94. Adepoju AO, Amusa MO, Ugwu OPC. Ethnopharmacological survey on medicinal plants utilization in Freetown, Sierra Leone. *RPS Pharm Pharmacol Rep.* 2023;2:1–13.
95. Ugwu OPC, Alum EU, Okon MB. The role of environmental DNA (eDNA) in biodiversity conservation. *Res Output J Biol Appl Sci.* 2024;3(1):79–83.
96. Alum EU, Izah SC, Uti DE, Ugwu OPC, Betiang PA, Basajja M, et al. Synergistic phytochemicals in multi-target drug discovery for complex diseases: a narrative review. *Phytomedicine Plus.* 2026:100995.

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